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# PRODUCTION TECHNOLOGY, NUTRITIONAL VALUE, AND THERAPEUTIC BENEFITS OF DRAGON FRUIT (HYLOCEREUS SPP.): A REVIEW

Ashok Kumar<sup>1</sup>, Jagdish Grover<sup>2</sup>, S.R. Singh<sup>3</sup>, M.C. Yadav<sup>4</sup>, Vijay Kumar Yadav<sup>5</sup>, Pooja Yadav<sup>6</sup>, Arvind Kumar Channi<sup>7</sup>, Navneet Kumar<sup>8</sup>, Joe Christy N<sup>9</sup> and Sanjay Kumar<sup>10</sup>

<sup>1</sup>Professor Horticulture-Fruit Science, Guru Kashi University, Bhatinda-161302, Punjab, India.

<sup>2</sup>Director Extension, Guru Kashi University, Bhatinda-161302, Punjab, India.

<sup>3</sup>Principal Scientist, Indian Institute of Sugarcane Research, Lucknow, Uttar Pradesh, India

<sup>4</sup>Principal Scientist, NBPGR (ICAR), Pusa Campus, New Delhi-110012, India.

<sup>5</sup>Professor Genetics & Plant Breeding, CSAUA&T- Kanpur, U.P., India.

<sup>6</sup>Assistant Professor, Agrometeorology, Guru Kashi University, Bhatinda-161302, Punjab, India.

<sup>7</sup>Associate Professor, Department of Mechenical Engineering, Guru Kashi University, Bhatinda-161302, Punjab, India. 
<sup>8</sup>Assistant Professor, Department of Mechenical Engineering, Guru Kashi University, Bhatinda-161302, Punjab, India. 
<sup>9</sup>Professor, Faculty of Management and Commerce (FMC), Guru Kashi University, Bhatinda-161302, Punjab, India.

 $^{10}\mbox{Associate Professor Agronomy, Guru Kashi University, Bhatinda-161302, Punjab, India.}$ 

\*Corresponding Author: Prof. (Dr) Ashok Kumar

Professor Horticulture-Fruit Science, Guru Kashi University, Talwandi Sabo, Bhatinda, Punjab, India Email: yadavakdr@gmail.com, drashokkumaryadav17203gku.ac.in

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ABSTRACT

Dragon fruit or Pitaya (Hylocereus spp.) a member of Cactaceae family is gaining popularity in the market for consumption of fresh fruit due to its highly attractive fruit color with nutritional and immense bioactive potential that reduces blood sugar level, cholesterol, prevention of liver injuries, cancer, and so forth. In commercial market valorized products of DFs are available. Additionally, dragon fruit extracts have hepatoprotective, cardiovascular, and prebiotic efficacy. Dragon fruit, which is native to northern South America and Mexico, has become a significant crop in tropical and subtropical regions worldwide, including Vietnam, China, and Australia. Although it originated in Central America, it is now grown all over the world, particularly in subtropical regions. It is a quickly expanding climbing plant that needs a vertical support pole to grow on before a ring fall like an umbrella. The fruit (Hylocereus spp.) is rich in various bioactive phytochemical compounds, including phenolic acids, flavonoids, and pigments such as betalains and anthocyanins, which contribute to its antioxidant, anti-inflammatory, and anti-microbial properties. A review of global industry standards reveled that Australian standard could incorporate classifications of dragon fruits, including external factors like appearance, size, and defect tolerance. Future research is needed to prioritize understanding of the impact of cultivation practices and environmental factors on the bioactive composition of dragon fruit, enabling the development of best practices for growers. The advancement of analytical methods for quantifying bioactive compounds will provide deeper insights into their health benefits and support the establishment of bioactive-oriented industry standards. Moreover, investigations of post-harvest handling and processing techniques could optimize the preservation of these valuable compounds, enhancing dragon fruit's role as a functional food.

Keywords: Dragon Fruit, Nutritional Values, therapeutic benefits, Australian markets, Industry standard.

#### 1. Introduction

Dragon fruit (Hylocerus undatus), introduced as a new crop in low rainfall zone and rocky barren land, is gaining rapid popularity amongst farmers. It is a nutritious fruit with a variety of uses as pulp, which constitute 70-80% of the ripe fruit. Dragon fruit is mainly available in three variants viz., red skin with white pulp (Hylocereus undatus), red skin with red pulp (Hylocereus monacanthus previously known as H. polyrhizus) and yellow skin with white pulp (Hylocereus megalanthus

previously known as Selenicereus megalanthus). The redfleshed varieties of dragon fruit are relatively rich in antioxidants. It is known to prevent colon cancer, diabetes and neutralizes toxic substances such as heavy metals; reduce cholesterol and high blood pressure. It is rich in vitamin C, phosphorus and calcium. The flavour of the fruit resembles to kiwi fruit. Fruits are low in fat and rich in minerals with the optimum Brix value of 15-180Bx. Dragon fruit (DF) is also known as Pitaya or Pitahaya, growing on the climbing vine-like epiphytic cacti plant species of genus Hylocereus and Selenicereus belongings to family Cactaceae and sub family Cactoideae (Le et al., 2021). The genus Hylocereus and Selenicereus are indigenous to the forest of Mexico, northern South America, and Central America (Mizrahi et al., 1997). The species of genus Selenicereus i.e Selenicereus megalanthus (Schum.) Britt., and Rose is a majorly significant species, also known as Yellow Pitaya or Pitaya amarilla, wildly grown in Nicaragua, Colombia, and Ecuador. Selenicereus megalanthus has also been mentioned as Hylocereus megalanthus, it does not have mutual morphological characteristics as Selenicereus species. The spiny structure  $(1-2\Box cm)$  at the outer surface of the fruit can differentiate the actual fruit of S. megalanthus (Yellow Pitaya) from H. megalanthus. It looks like other species of the genus Hylocereus but color of the peel is yellow (Mizrahi, 2014).

#### (i) World dragon fruit production

Being a native of Southern Mexico, Guatemala and Costa Rica, dragon fruit was introduced during 1990 for its commercial cultivation in south Asian tropical countries. At present, significant production and expansion of fruit is occurring in many countries viz., Vietnam, China, Mexico, Colombia, Nicaragua, Ecuador, Thailand, Malaysia, Indonesia, Australia and United States. However, its production and marketing data is rarely available apart from its spontaneous expansion around world. Dragon fruit is classified as group of minor tropical fruit depending on market availability, production, planted areas and economics consideration (FAO, 2012; Liaotrakoon, 2013). Further, it is believed to be listed in FAO database under item name tropical freshness. Along other tropical fruits. Available evidences from individual countries and recent reports published by some of the private organisations suggesting the dragon fruit production increased significantly in past and current decade. At present world market contributes four types of dragon fruits: (i) red skin, white flesh (Hylocereus undatus), mainly from Vietnam and Thailand; (ii) red skin, red flesh (Hylocereus polyrhizus) come mainly from Israel and Malaysia; (iii) red skin, purple flesh (Hylocereus costaricencis) from Guatemala, Nicaragua, Ecuador, and Israel; and (iv) yellow skin, white flesh (Hylocereus (Selenicerus) megalanthus) from Colombia and Ecuado. The red-skin with white flesh, red-skin with red flesh, and red-skin with purple flesh and yellow-skin

with white flesh accounts approximately 94, 4.0, 1.5 and 0.5% shares in world market. Guesstimates can put on current world dragon fruit production to be more than 2.1 million tons (Mt) in an area of 1.12 lakh ha (2017–18). Vietnam, China, Indonesia, Thailand, Taiwan, Malaysia, Philippines, Cambodia, India and USA are the leading producers. Three major countries viz., Vietnam, China and Indonesia contribute more than 93% of dragon fruit production of the world (Fig. 2). The share of Vietnam alone is more than half (51.1%) of the world production over an area of 55, 419 ha with average productivity of 22-35 t/ha/year. The volume of dragon fruit produced in Vietnam is more than 1 Mt of worth US\$ 895.70 million (Chen and Paull, 2018). Dragon fruit cultivated in almost all provinces of Vietnam, but largely concentrated in Binh Thaun, Tein Giang and Long a region (Hein, 2018). China is second largest producer contributing 33.3% of world production of dragon fruit i.e., producing about 7, 00,000 tons dragon fruit of worth US\$ 397 million over growing areas of 40,000 ha with average productivity of 17.5 t/ha/year (Table 1; Fig. 2; MZMC, 2020). Among major provinces and regions of China, Guangxi has the largest growing area of 13,000 ha, followed by Guangdong with 10,000 ha and Hainan and Guizhou, each with about 6,600 ha (MZMC, 2020). Indonesia is third largest producer of dragon fruit contributing 10.6% of world production and produces annually about 2.22 lakh tons dragon fruit over an area of 8,491 ha. Dragon fruit is mainly cultivated in Java, Kalimantan, Sumatra and Bali Island of Indonesia and its average productivity varies from 15-30 t/ha/year. After 2014, more than 700 ha of dragon fruit orchards in East Kalimantan and West Sumatra provinces became unproductive due to severe damage of stem canker disease. Taiwan and Thailand contribute approximately 2.3 and 1.2% in world production. The production area in Taiwan rising steadily from 380 ha in 1999 to 2,490 ha in 2017 owing to farmers fetches stable prices. In Thailand, the main planting areas are located in the lowlands of Central Plains around Bangkok. Hence, orchards with raised furrow and ditch system are surrounded by soil barrier to prevent flooding during rainy season (Saradhuldhat, 2018). The production and area under dragon fruit is also increasing in other south Asian tropic countries viz., Malaysia and Philippines. The commercial dragon fruit production is also introduced in USA, Cambodia, Australia and South Africa in recent decades (0.5%). During 2017–18, dragon fruit production of India is about 4,200 tons, which is only 0.2% of the world production.

## (ii) Major dragon fruit supplier countries

World major supplier countries of dragon fruit can be broadly divided into following three main hubs. 1. Asia: Vietnam, China, Thailand, Taiwan, Indonesia, Malaysia, Philippines, Cambodia, India and Sri Lanka. 2. Middle East and Europe: Israel, Switzerland and EU. 3. America: Mexico, Colombia, Ecuador, Guatemala and

Costa Rica. At present, Vietnam is the largest dragon fruit supplier in the world, accounting the highest share in Asia, Europe and sometimes in the United States. Vietnam dragon fruit has been a long trademark of Asian Americans. Particularly in Asia; Vietnam is a leading supplier to the Chinese market. Other market viz. Hong Kong, Taiwan, Thailand and Malaysia are also trying to establish their trade in Asian markets. According to American consumer's website, Vietnam's red dragon fruit is generally bigger than other types of dragon fruit, with a nice and impressive appearance, but it is light in colour not as crispy and sweet as yellow dragon fruit. In contrast, the red dragon fruit of Vietnam, although not highly evaluated in form, but is rated to have superior flavour compared to other red dragon fruit. Israel and Thailand are known major dragon fruit suppliers for European countries. In the US market; Mexico, Central and South America are the biggest competitors to Asian dragon fruit suppliers owing to geographical advantage. In Central America, Nicaragua are major producers and suppliers of red dragon fruit to USA, Canada, Europe and Japan. Colombia is the leading supplier of yellow dragon fruits and highly valued for their taste and form in the US market, as they are the sweetest and most vibrant of the dragon fruit varieties. In addition, this dragon fruit has seasonal advantages to capture the markets when other types of dragon fruits, do not arrive as it is usually harvested in November to February every year. Ecuador produces both yellow and red dragon fruit. Guatemala, El Salvador, Honduras, Costa Rica produce dragon fruit in small scale particularly in the garden (dragonfruit.net.vn).

## 2. Botanical characteristics of dragon fruit:

Some of the botanical characteristics of the plant are mentioned below. • Hylocereus undatus is a climbing vine cactus species of the family Cactaceae. It is is a fast growing, epiphytic or xerophytic. • It is one of 15 accepted Hylocereus species. While many of these have ornamental value because of their flowers, only five are important as fruit producers. • Common names such as 'dragon fruit', 'pitaya' and 'pitahaya' are generic terms, which include several species of columnar and climbing cacti. They are often applied to species other than H. undatus too and usually refer to the fruits rather than the plant. • It is considered native to southern Mexico, the Pacific coast of Guatemala, Costa Rica, and El Salvador. It is now commercially cultivated and widely distributed throughout the tropics and some temperate regions. • Stems are triangular, 3-sided, although sometimes 4- or 5sided, green, fleshy, and jointed, many branched. Each stem segment has 3 flat wavy ribs and corneous margins may be spineless or have 1-3 small spines. • Stems scandent, creeping, sprawling or clambering, up to 10 m long. Aerial roots, which are able to absorb water, are produced on the underside of stems and provide anchorage for stems on vertical surfaces. • Flowers are

25-30 cm long, 15-17 cm wide, nocturnal, scented and hermaphroditic; however, some cultivars are self-compatible. Flowers are typically white in colour and bell shaped, stamens and lobed stigmas are cream coloured. • Fruit is a fleshy berry, oblong to ovoid, up to 6-12 cm long, 4-9 cm thick, red with large bracteoles, pulp white, edible, embedded with many small black seeds. • Average fruit weight is 350-400 g, although may weigh up to 900 g.

(i) Flower Biology: Dragon Fruit Flower is being pollinated with the help of Asian Honey Bee August 24, 2021 607 1 Singapore Geographic, Singapore Nature, Singapore Nature Photography Pollination is the transfer of pollen from a male part of a plant to a female part of a plant, later enabling fertilisation and the production of seeds, most often by an animal or by wind. Pollinating agents are animals such as insects, birds, and bats; water; wind; and even plants themselves A pollinator is an animal that moves pollen from the male anther of a flower to the female stigma of a flower. This helps to bring about fertilization of the ovules in the flower by the male gametes from the pollen grains. Insect pollinators include bees, (honey bees, solitary species, and pollen wasps (Masarinae); bumblebees); fliesincluding beeflies, hoverflies, blowflies and mosquitoes; lepidopterans, both butterflies and moths; and flower beetles. Vertebrates, mainly bats and birds, but also some non bat mammals (monkeys, lemurs, possums, rodents) and some lizards pollinate certain plants. Among the pollinating birds are hummingbirds, honeyeaters and sunbirds with long beaks; they pollinate a number of deep-throated flowers. Humans may also carry out artificial pollination. The most recognized pollinators are the various species of bees, which are plainly adapted to pollination. Bees typically are fuzzy and carry an electrostatic charge. Both features help pollen grains adhere to their bodies, but they also have specialized pollen-carrying structures; in most bees, this takes the form of a structure known as the scopa, which is on the hind legs of most bees, and/or the lower abdomen (e.g., of megachilid bees), made up of thick, plumose setae. Honey bees, bumblebees, and their relatives do not have a scopa, but the hind leg is modified into a structure called the corbicula (also known as the "pollen basket"). Most bees gather nectar, a concentrated energy source, and pollen, which is high protein food, to nurture their young, and inadvertently transfer some among the flowers as they are working. Euglossine bees pollinate orchids, but these are male bees collecting floral scents rather than females gathering nectar or pollen. Female orchid bees act as pollinators, but of flowers other than orchids. Eusocial bees such as honey bees need an abundant and steady pollen source to multiply. Honey bees travel from flower to flower, collecting nectar (later converted to honey), and pollen grains. The bee collects the pollen by rubbing against the anthers. The pollen collects on the hind legs, in a structure referred to as a "pollen basket". As the bee flies from flower to flower, some of the pollen grains are transferred onto the stigma of other flowers. Nectar provides the energy for bee nutrition; pollen provides the protein.



**Fig. 1:** Flower of Dragon Fruits (Side View)

When bees are rearing large quantities of brood (beekeepers say hives are "building"), bees deliberately gather pollen to meet the nutritional needs of the brood. Dragon Fruit Flower also called Pitaya is an exotic fruit cactus with colorful leaves, flowers, and fruits. Dragon fruit cactus bloom during the night and hence called the queen of the night. It opens up at dusk and stays abloom till midnight. Before flowering, the flower buds can grow up to 10" inches long. Flowering begins in summer, usually at the beginning of June, soon followed by the formation of fruits. It takes at least 13 days for the flowers to form and 50 days for the fruits to become mature. Dragon Fruit flowers are night bloomers and not capable of self-pollination, they are highly dependent on bats, moths or bees to do the job. A dragon fruit plant requires the pollen of a second dragon fruit plant of a different variety in order to set fruit. Here is the video of Asian honey bees (Apis cerana) with a dragon fruit flower in the early morning, there were 3 flowers, and bees were visiting these 3 flowers.

The most popular, largely produced, and highly money-making species of genus Hylocereus is Hylocereus undatus (Haworth) Britt. and Rose. This species was brought by the French cleric to the Vietnam in 19th century and now the country has become the largest exporter of DF. There are other known species such as Hylocereus polyrhizus (Weber) Britt. and Rose, Hylocereus costaricensis (Weber) Britt. and Rose, Hylocereus megalanthus, Hylocereus ocamponis, Hylocereus purpusii, Hylocereus trigonus, and so forth (Mizrahi et al., 1997; Perween et al., 2018) these are oval, oblong, or tuber in shape and distinguished on the basis of the size of the fruit, color of peel and color of pulpy flesh which consists of many black colored tiny seeds.



Fig. 2: Flower of Dragon Fruits (Front View)



Fig. 3: Flower of Dragon Fruits (On plant View). Sources: By Mary H. Dyer, last updated December 15, 2022



**Fig. 4:** Flower of Dragon Fruits (View of Flowers). *SOURCES: Brand: The Entacloo* 



**Fig. 5:** Flower Biology of Dragon Fruits with Pollination. *Sources:* https://en.wikipedia.org/wiki/Pollinato



Fig. 6: Pictorial display of Hylocereus species (A) Hylocereus polyrhizus, (B) H. undatus, (C) Hylocereus costaricensis, (D) Hylocereus megalanthus, (E) Selenicereus megalanthus (without spines), (F) S. megalanthus (with spines), (G) Hylocereus purpusii, and (H) Hylocereus trigonus. Sources: Chen, Siyuan, Xu, Cheng-Yuan, Mazhar, Muhammad, Naiker, Mani, 2024.

The morphological characteristics of different varieties are shown in Table 1 (Bellec & Vaillant, 2011; Perween *et al.*, 2018). There are many international common names for DF in different languages and many local names in different countries (Tables 2 and 3) (CABI, 2020; Mizrahi *et al.*, 1997).

Table 1: Physical characteristic of Hylocereus species.

Species	Size (cm)	Weight (g)	Physical appearance		
Hylocereus undatus	15–22	300–800	Rosy red colored peel with white flesh, green at the top		
Hylocereus polyrhizus	10–12	130–350	Red colored peel with red flesh, green at the top		
Hylocereus purpusii	10–15	150–400	Light or dark red colored peel with red flesh, green at the top		
Hylocereu scostaricensis	10–15	250–600	Red colored peel with red-purple colored flesh		
Hylocereu strigonus	7–9	120–250	White colored flesh		
Hylocereus megalanthus	7–9	120–250	Yellow colored peel with white colored flesh		

Source: Bellec and Vaillant (2011); Perween et al. (2018).

**Table 2 :** Language wise names for *Hylocereus species* spoken in worldwide.

Languages	International common names		
English	Belle of the night; Night-blooming cereus;		
	Pitahaya; Red pitahaya; Queen of the night;		
	Strawberry pear		
Portuguese	Cardo-ananaz; Cato-barse		
French	Fruit du dragon; Pitahaya rouge; Pitaya		
Spanish	Pitahaya blanca; Pitahaya dulce; Pitahaya roja		
Chinese	Liang tian chi		

Source: CABI (2020).

**Table 3 :** Local names of *Hylocereus species*.

Country	Local names		
Vietnam	Thanh long		
Mexico	Junco tapatio; Pitahaya de cardo; Pitahaya orejona		
Germany	Distelbirne; Drachenfrucht; Konigin der nacht;		
	Rotepitahaya		
Brazil	Pitaya vermelha de polpabranca		
USA/Hawaii	Paninniokapunahou; Papipipua		
Sweden	Rod pitahaya; Skogskaktus		
Puerto Rico	Flor de caliz; pitajava		
Cuba	Flor de cáliz; Pitahaya		

Source: CABI (2020); Mizrahi et al. (1997).

H. undatus (White flesh dragon fruit [WFDF]) and H. polyrhizus (Red flesh dragon fruit [RFDF]), are the most popular and commercialized species grown in various parts of the world. DF comes under the category of nonclimacteric fruits (Zee et al., 2004). It grows in the tropical and subtropical areas of the world like Southern and Central America and South-East Asia within the temperature range 25-29°C. It can resist at temperature 38-40°C and minimum at 0°C for a short time period. Generally, 2-4, DF plants climb on a single standing support system (1.5 to 2 m) made up of concrete, wood, and so forth. as shown in Figure 2 (Liaotrakoon, 2013). Normally a 20–30 cm long old plant (mother plant) stem is used for planting of new plants of DF (at the end of the fruiting season). For the rooting purpose, these stems are put into the poly bags which contain sand, soil, and farm backyard muck (carbonized rice husk and coconut coir can also be used) in the ratio 1:1:1 and, then held for rooting. Generally, rooting occurs 5-6 months but with application of IBA (Indole-3-butyric acid), a synthetic hormone of auxin family, quick rooting occurs within 1 month (Frick & Strader, 2018). To avoid disease attack, it is conditioned with fungicides. It takes 8-9 months for flowering (25–30 cm in length) in the entire sunny arena. Planting through seeds of DF produces a plant with a thin stem and many variations. Flowers of the plant blossom and spread fragrance during the night only. The total time period required for the overall process (i.e., from flower to fruit) is approximately 4-5 months; it starts from the month of April or May and continues till September or October. H. undatus species takes 50 days for the completion of the maturation cycle to produce a complete fruit whereas S. megalanthus takes 150 days (Mizrahi et al., 1997). During maturation, the color of fruit changes from green to red. Regularity in the supply of water to the plant is necessary to get high-quality fruit. In India, highly favorable defrost and dry weather for the cultivation of DF is available in North-East, South, and West region. In India, the wholesale rate of DF in the market is in the range of INR 80-120 per kg but during the off-season rate increases upto INR 150-250 per kg (Karunakaran et al., 2019; Saradhuldhat. 2019). DF could be stored for 25–30 days at refrigeration temperature (4°C), whereas <10 days at room temperature (Liaotrakoon. 2013). According to Paull (2016), 10°C is best storage conditions for DF storage (upto 14 days) in respect of overall quality, because at 4°C chilling injury, a change in the physical structure of peel and change in taste of fruit occurs. DF should be harvested after 30–35 days of flowering.



Fig. 7: Inter Culture in Dragon Fruits Orchards, Sources: Guru Kashi University, Bathinda, Punjab, India



Fig 8: Dragon Fruit Cultivation. Sources: Ministry of Agriculture & Farmers Welfare, 13 MAR 2023 8:03PM by PIB Del

Majorly DF is produced in Vietnam and this country is the largest exporter in the world as well. Area cover under the production of DF in Vietnam was 5512 ha in the year 2000 but with the rapidity of development in the year 2018 it was near about >55,000 ha and the major producing cities are BinhThuan (Highest producer) followed by Long An, Tien Giang, Dong Nai, Tay Ninh, and so forth. These cities grow a higher amount of white flesh DF (95%) and left (5%) is red flesh DF. About 80%–85% of the total produced DF is exported from Vietnam (Hien., 2019). According to the report of the department of crop production Vietnamese Ministry of agriculture and rural development, the country had exported a heap of the DFs which was accounting for the highest percentage at 33.9% among all other fruits.

Whereas about India, it was newly introduced fruit after 1990s and area underneath an expansion of agriculture production is in developing state. Presently some parts of Indian states (<400 ha) such as Maharashtra, Karnataka, West Bengal, Tamil Nadu,

Kerala, Gujarat, Andaman, and Nicobar Islands, Orissa, and Andhra Pradesh are cultivating DF, but production is not fulfilling the market demand of India. So, largely DF is imported in India from countries like China, Vietnam, Malaysia, Sri Lanka, and Thailand (Karunakaran *et al.*, 2019). Other countries producing DF includes Malaysia, Mexico, Philippines, Indonesia, and so forth. Majorly DF is imported by European Union (57%) followed by USA (17%), Germany (14%), France (12%), Netherlands (11%), Russia (10%). United Kingdom (9%), and Canada (8%) (Mercado-silva., 2018). The country-wise production of DF in the year 2014 is given in Table 4 (Mercado-silva., 2018).

#### (ii) Nutritional Importance

DF comes in the category of functional foods due to its highly nutritious and nutraceutical properties. Nutritionally, it consists of vitamins such as vitamin C, vitamin A, and niacin, carbohydrates, proteins, fats, minerals such as potassium, sodium, calcium, phosphorus, zinc, magnesium, and iron, and fibers.



**Fig. 10:** Dragon fruit plantation system with fruitings. *Sources: Agribusiness (Dragon Fruit Farming)* 

The nutritional value of DF is given in Table 5 (Ali Jaafar *et al.*, 2009; Liaotrakoon, 2013; To & Duc, 2000; USFDA, 2022). DF plays a beneficial role for health-conscious people because of the very low content of fat, lots of minerals, vitamins, higher number of dietary fibers.

**Table 4:** Datasheet of DF production in year 2014 (country wise).

Producer country	Production (tones)	Output (tones/ha)
Vietnam	6,02,680	40–45
China	2,00,000	14
Indonesia	36,000	24-30
Taiwan	27,654	20-30
Malaysia	8577	19-30
Nicaragua	6160	6–9
Mexico	2496	4–6
Myanmar	2000	20
Philippines	1574	6
Israel	1000	25-40

Source: Mercado-silva (2018).

**Table 5 :** Nutritional composition of DF with % of RDA cover by DF.

<b>Nutritional composition</b>	WFDF	RFDF	RDA (%)
Moisture (%)	85.3	82.5-83	-
Total carbohydrates (g)	11.2	11.2	4
Fructose	1.9	1.8-3.2	-
Glucose	5.5	4.7–5.7	-
Fat	0.1	0.21-0.61	0.13-0.78
Sorbitol	0.33	-	-
Protein	1.1	0.159-0.229	0.3-2.2
Crude fiber	1.34	0.7-0.9	-
Vitamin C (mg/100 g)	3	8-9	3–10
Vitamin A	0.0111	-	1.2
Niacin	2.8	-	17.5
Potassium	272	272-328.4	5.8–7
Magnesium	38.9	31.3–38.9	7.4–9.2
Phosphorus	27.5	27.5–36.1	2.2-2.8
Calcium	10.2	2.3-10.2	0.17-0.2
Iron	3.37	0.6-3.4	3.3-18.8
Sodium	8.9	7.3–8.9	0.32-0.38
Zinc	0.35	-	3.2
Energy (KJ)	130	283	_

Source: Jaafar et al. (2009), Liaotrakoon (2013), To and Duc (2000), USFDA (2022).

**Abbreviations:** RDA, recommended dietary allowance; RFDF, red flesh dragon fruit; WFDF, white flesh dragon fruit.

#### (iii) Physico-chemical Properties

Compounds like phenolic and their subdivisional groups, antioxidants, flavonoids, carotenoids, anthocyanins, essential oils, colorants, polyphenols, phytosterols, and so forthcome under bioactive compounds. These types of bioactive compounds are found in DF with a higher amount which is a boon for consumers. Bioactive compounds present in DF peel are more than flesh. The amount of various bioactive compounds in DF is given Table 6 (Wu *et al.*, 2006).

**Table 6:** Comparison of bioactive compounds found in red flesh dragon fruit and peel.

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Bioactive compounds		DF
	flesh	peel
Total phenolic content (mg GAE/100 g)	42.4	39.7
Total flavonoid content (mg CE/100 g)	7.21	8.33
Betacyanins (mg BE/100 g)	10.3	13.8
Antioxidant activity		
DPPH radical scavenging activity (µmol VCE/g)	22.4	118
ABTS radical scavenging activity (µmol TEAC/ g)	28.3	175

Source: Wu et al. (2006).

**Abbreviations:** BE, betanin equivalent; CE, catechin equivalent; DF, dragon fruit; GAE, gallic acid equivalent; TEAC, trolox equivalents antioxidant capacity; VCE, vitamin C equivalent.

The pink or red color of the peel and flesh of DF is only due to the presence of betacyanins. Betacyanin is a red and violet colored compound from pigment that is, betalains that is water-soluble and contains nitrogen molecules (Esatbeyoglu *et al.*, 2015). Betacyanins are

classified into three groups such as Betanin (Phyllocactin, 2-apiosyl-phyllocactin, 2-betanin, hylocerenin), Amaranthin, and Gomphrenin (Gomphrenin class I, II, III, and betaninidin 6-O-sophorosides derives) (Strack et al., 2003). Out of these, the members of betacyanins are found in DF are betanin, isobetanin, phyllocactin, isophyllocactin, hylocerenin, and isohylocerenin (Stintzing & Carle, 2007). The chemical structures of different betacyanins are given in Figure 3 (Liu et al., 2019; Stintzing & Carle, 2007). The tiny black seeds which are present in DF flesh are also important in terms of oil. DF seed oil, comprised of a higher amount of essential fatty acid that is, linoleic acid and linolenic acid covers approximately 56% of total seed oil followed by monounsaturated (oleic acid, palmitoleic acid, etc) and saturated fatty acids (palmitic acid, stearic acid, etc). WFDF seed contains little more essential fatty acids in comparison to RFDF as shown in Table 7. Along with fatty acids, DF seed oil also contains triacylglycerol (majorly tri or di-unsaturated triacylglycerol and somewhat monounsaturated triacylglycerol) (Liaotrakoon, Clercq et al., 2013).

Tocopherol concentration is also found in DF seed oil such as 407.26 mg/kg in WFDF seed oil and 656.89 mg/kg in RFDF seed oil out of which 71%-73% is occupied by  $\alpha$ -tocopherol followed by  $\gamma$  -tocopherol and (18%-22%)remaining δ-tocopherol with (Liaotrakoon Van Buggenhout, et al., 2013) and is reported nearly similar to sesame oil (454-700 mg/kg) (Aued-pimentel et al., 2006), comparably more than olive oil (160-378 mg/kg) (Krichene et al., 2010), corn oil (115.4 mg/kg) (Lemcke-noroja et al., 2001), grape seed oil (143-358 mg/kg)(Baydar et al., 2007), strawberry seed oil (280 mg/kg) (Van Hoed et al., 2009), linola seed oil (20.8 mg/kg) (Lemcke-noroja et al., 2001), and cranberry seed oil (139 mg/kg) whereas less than red raspberry seed oil (2106 mg/kg) and blackberry seed oil (1369 mg/kg) (Van Hoed et al., 2009). Comparative status of various bioactive components in seed oils of red flesh and white flesh DF is given in Table 7 (Liaotrakoon, Clercq et al., 2013).

**Table 7:** Bioactive compounds of WFDF and RFDF seed oil

Bioactive compounds	RFDF seed oil	WFDF seed oil
Linoleic acid (%)	45.21	55.43
Linolenic acid	0.15	0.21
Hexadecadienoic acid	0.11	0.15
Archidonic acid	0.04	0.07
Eicosatrienoic acid	0.18	0.07
Total essential fatty acids	45.37	55.64
Total polyunsaturated fatty acids	45.7	55.94
Oleic acid	23.61	18.67
Palmitoleic acid	1.04	0.79
Gadoleic acid	0.24	0.16
Erucic acid	0.07	0.05
Total monounsaturated fatty acids	24.96	19.68
Palmitic acid	18.39	14.95
Stearic acid	8.05	6.93

Arachidic acid	1.04	0.95
Behenic acid	1.12	0.83
Lignoceric acid	0.41	0.44
Myristic acid	1.21	0.98
Total saturated fatty acids	29.26	24.34
Protocatechuic acid (mg/100 g)	0.93	0.93
p-Coumaric acid	0.78	0.79
p-Hydroxybenzoic acid	0.66	0.72
Vanillic acid	0.64	0.7
Gallic acid	0.25	0.2
Syringic acid	0.08	0.21
Caffeic acid	0.08	0.71
Total phenolic content of oil	3.42	4.26
Beta-sitosterol	676	548
Campesterol	252	198
Stigmasterol	106	72
Cholesterol	6	7
Total phytosterol content	1040	825
Total tocopherol content	656.89	407.26

Source: Liaotrakoon, Clercq et al., 2013.

**Note:** Bold values indicate the total values of the respective category of nutrients.

**Abbreviations:** RFDF, red flesh dragon fruit; WFDF, white flesh dragon fruit.

## 3. Cultural Practices

- (i) Climate and Soil: Dragon fruit has potential to grow in various agro-climatic conditions of tropical and subtropical region. Dragon fruit plant prefers a dry tropical climate with an average temperature of 20-29°C, but can withstand temperatures of 38-40°C and as low as 0°C for short periods. The rainfall requirement of dragon fruit is 1145- 2540 mm/year. Dragon fruit plants prefer sandy loam with high organic matter and grow well in soil having good drainage. A soil pH 5.5-6.5 is optimum for growth.
- (ii) Varieties: Red Pithaya, Pitaya Roja, Pitaya Amarilla, Yellow pitahaya, Alice, American beauty, Bloody mary, Cosmic Charlie, Costarican sunset, Dark star, David bowie, Delight Propagation: Dragon fruit plants can easily multiply through stem cutting.
- (iii) Planting: Generally, 20-25cm long stem cuttings are used for planting during July-August. Dragon fruit cultivation prefers full sunlight open area is very suitable for planting. Single post vertical height of pole 1.5 m to 2 m at which point they are allowed to branch and hang down. The Dragon fruit may be planted near the poles to enable them to climb easily. Number of plants per pole may be 2 to 4 plants depending on the climatic condition. Spacing: Generally, in single post system planting is done at 3m x 3m distance in dragon fruit.
- (iv) Manures and Fertilizers: Application of FYM @15-20kg/plant/year and a fertilizer dose of N 450g:  $P_2O_5$  350g:  $K_2O$  300g/plant/year perform best result for dragon fruit yield and quality. After care: Regularly prune the plants to obtain an open and manageable umbrella shape canopy which will induce new shoots for the next

cropping season. The well grown vine may produce 30 to 50 branches in one year and may be more than 100 branches in-four years.

Weed Management: Weeding should be done regularly. Weed control is an important operation in dragon fruit cultivation and the use of weed mat efficiently reduced the weeds growth and also aids in soil moisture conservation. Water Management: Dragon fruit root system is shallow and distributed in 15 to 30 cm depth. Hence irrigation should be insured to provide sufficient water during dry season. Drip irrigation found beneficial for better yield and growth. Approximately 2-4 litres of water weekly twice per plant is sufficient during the summer/dry days.

- (v) Harvesting and Yield: The plant start yielding after 12-15 months from the date of planting. The plants yield the fruits in the months between June to September, and harvest could be done three to four times in a month, the fruit weight ranged between 300-800g, and the average yield from the single post is realized about 30 to 35 kgs from the three years old planting. On an average, dragon fruit yield is 8-10 tonnes/ha/year.
- (vi) Post Harvest Management: Dragon fruits are graded by size and colour. Dragon fruits can be stored for 4 weeks at 6-100C and 85-90% relative humidity.

## (vii) Planting

Dragon fruit cultivation prefers full sunlight open area for planting. Generally, in single post system planting is done at 3x3 m distance. Single post vertical height of pole 1.5 m to 2 m at which point they are allowed to branch and hang down. The Dragon fruit may be planted near the poles to enable them to climb easily. Number of plants per pole may be 2 to 4 plants depending on the climatic condition. Lateral shoots must be limited and 2-3 main stems are allowed to grow. It is important to arrange round metal/concrete frame to maintain balanced shrub. Cost effective structures using iron poles and tyres as base structure are also being used. In high rainfall areas, concrete poles are used as trellises. The concrete pillars are supported by a square structure in the top to train the vine for bearing purpose. To combat soil erosion, the concrete square structures are used as base for maintaining the media that supports the growth of the vine. The planting season is generally summer monsoon (June-August). Fruiting occurs in July-October in a 6-8 flushes of market quality fruits.

## (viii) Training

The Dragon fruit plants are fast growing vines and produce thicker dense of branches during the initial stage. The lateral buds and branches should be pruned to grow towards stands. Once vines reach up to the top of the stands the branches are then allowed to grow. The removal of tip of main stem is done to allow growth of new shoots to grow laterally and climb at the ring to form

an umbrella like structure of vines where flowers will emanate and develop into fruits which would induce lateral branching. This pruning referred as structural pruning or making a structure on the trellis. The well grown vine may produce 30 to 50 branches in one year and may be more than 100 branches in-four years.

#### (ix) Nutrient management

Dragon fruit plant root system is superficial and can rapidly assimilate even the smallest quantity of nutrients. The recommended dose of fertiliser application varies with soil type and location of plantation. In general, 10–15 kg FYM or organic manure and 100 g SSP/plant is compulsory at the time of plantation of dragon fruit. About 300 g N, 200 g P and 200 g K is essential per plant each year for the initial two years.

The mature plant should be given 540 g N, 720 g P and 300 g K in four equal split doses with interval of 3 months. The nutrients are supplied in four split doses to each pillar having four plants @ 10, 10 and 30% of total, before flowering, 20, 40 and 25% at fruit set, 30, 20 and 30% at harvest and finally 40, 30 and 15% of total NPK after two months of harvest. A combination of organic manure with neem cake and 100g of complete fertilizer (19-19-19) is applied every three to four months.

#### (x) Intercultural operations

Weed control is an important operation in dragon fruit cultivation and the use of weed mat efficiently reduced the weeds growth and also aids in soil moisture conservation. Regularly prune the plants to obtain an open and manageable umbrella shape canopy which will induce new shoots for the next cropping season.

#### (xi) Irrigation

Regular irrigation is important, because it enables the plant to build sufficient reserves not only to flower at the most favorable time but also to ensure the development of the fruits. Local drip irrigation found beneficial for better yield and growth. Irrigation by flooding is not recommended as it wastes water and increases work of weeding. Approximately 2-4 litres of water weekly twice per plant is sufficient during the summer/dry days.

## 4. Bioactive Potential

The naturally occurring essential and nonessential components being part of the human diet in small quantities are known as bioactive compounds. Bioactive compounds of DF and their potential have been explored intensively because of their beneficial effects on high cholesterol, cardiovascular disease, and cancer. Different bioactive compounds have been found in DF as well as in RFDF such as betacyanins, fatty acids, phytosterols and acetic acid and so forth. (Foong *et al.*, 2012). Depending upon their chemical structure and functions they are divided into different categories such as flavonoids,

pigments and so forth. Few fatty acids are also found to be part of DF such as stearic and oleic acid that revealed to have a beneficial effect against cholesterol levels. According to Craig et al., (2000) it was suggested that stearic acid present in DF and RFDF decreases cholesterol absorption by decreasing plasma cholesterol. Studies also revealed that oleic acid also reduces cholesterol levels and therefore minimizes the risk of cardiovascular diseases. Different techniques such as fermentation are proved to increase the bioactive component such as fatty acids and phytosterols in DF and RFDF (Fadahunsi and Sunni, 2010). DF has been examined for the anti-insulin resistant activity for 6 weeks in which results showed that DF reduced the insulin resistance as well as hypertriglyceridemia and atherosclerotic in fructose supplement induced in rats. It was also found that thermally treated DF and RFDF at 95°C for 30 min enhanced hyperinsulinemia while as DF and RFDF treated thermally at 105°C for 60 min given to rats had no significant effect on insulin resistance, and atherogenesis. Antioxidant activity and content of soluble fiber contents are also attributed to the maintenance of diabetes (Omidizadeh et al., 2014). RFDF contains bioactive compounds in high contents and is known as a good source of polyphenols that can be incorporated into the diet, these phenolic compounds have a beneficial effect against cancers and possess strong antioxidant activity and also have potent antimicrobial properties (Foong et al., 2012). It has been suggested that flavonoids and phenolic acids were the foremost contributors to antioxidant activity and anti-LDL peroxidation. It has been studied that there was a decrease in total cholesterol, triglyceride, and HDL levels after giving DF peel as well as RFDF powder to male mice. RFDF contains high content of minerals and crude fiber particularly phenols (Setiawan et al., 2018). Different methods are used to measure or quantify these bioactive compounds present in DF and RFDF such as ethylenebenzothiozoline-6-sulfonic acid radical cation assay and ferric reducing antioxidant power assay are used to measure the antioxidant compounds. The ultrasonic-assisted extraction technique is also used to extract the antioxidant components (Ramli et al., 2014). DF extracts have been proved effective against both gram-negative, gram-positive bacteria and also showed less toxicity according to histopathological examination. DF extract cream has been also climates (Hossain et al., 2021) used for wound healing. (Mahdi et al., 2018). It was also proved that DF extracts reduce the levels of lactic acid and activity of creatinine kinase in rats treated with heavy physical exercise. Dose of 300 and 200 mg/kg were potential doses in reducing lactic acid and creatinine kinase (Harahap and Amelia, 2019). Therefore, DF was valuable in a multiplicity of applications in different pharmaceutical, biomedical fields, industrial applications. According to a new lifestyle of people in terms of food, they have been focusing on highly nutritious products, without chemical

additives, vegan, and so forth due to the awareness and consciousness towards health to boost the immune system. DF is reported to contain antioxidants, polyphenols, betacyanins, essential fatty acids, and sterols. Antioxidants like tocopherols, flavonoids, and polyphenols are useful for the prohibition of cellular components oxidation and total phenolic content for free radical scavenging activity. The antioxidants from the peel of DF showed prohibition in the growth of cancer cells that is, B16F10 melanoma (Wu et al., 2006), breast cancer cells (MCF-7) (Jayakumar & Kanthimathi, 2011), and hepatocellular carcinoma (HepG2) (Li et al., 2013) as compared to the flesh. The anticancer effect of DF is due to polyphenols which suppresses the activity of free nitric oxide (NO) radicals that inhibit the growth of cancerous cells (Jeong et al., 2010). Inhibition of MGC-803 growth was reported through stigma-4-en-3-one and Betasitosterol (Huo et al., 2014). Also, it showed hypocholesterolemic effect on lipids and reduces the abnormal amount of lipids found in the blood that is, it helps to reduce the total cholesterol concentration level, low-density lipoprotein, and triglycerides with the increment in the level of high-density lipoproteins which meliorate lipid present in the blood (Rokiah et al., 2009; Setiawan et al., 2018).

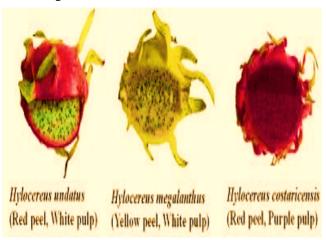
Betacyanins showed a magnificent effect against obesity which decreases the gain in weight of the body by decreasing the weight of the kidney, heart, and liver. Due to the consumption fatty liver disease. This problem can be reduced by the effect of betacyanins. Also, it reduces the exaggerated size of brown and white adipocytes (Song et al., 2016). Methanol extract of DF is very effective against carbon tetrachloride (CCl<sub>4</sub>) (responsible for creating toxicity in the liver that damages the liver) and enhances the effect of protecting from liver injury. Also, the effect of RFDF extract is comparable with silymarin used for the treatment of liver diseases (Mohammed et al., 2013). Dragon fruit (Hylocereus spp.), commonly referred to as pitaya, originates from Mexico and northern South America and has grown into a globally cultivated crop due to its adaptability to tropical and subtropical Initially valued for its vibrant appearance and exotic appeal, dragon fruit has gained significant attention for its rich contents of bioactive phytochemicals such as flavonoids, phenols, anthocyanins, and betalains, which contribute to its health benefits. These bioactive compounds are known for their antioxidant, antiinflammatory, anti-microbial, and anti-cancer properties, further elevating dragon fruit's value as a functional food (Kumar et al., 2018). The commercial cultivation of dragon fruit has spread across key producing countries like Vietnam, China, and Indonesia, where favorable climatic conditions support its growth and large-scale production (Paull, R.E.; Chen, N.J., 2019). In contrast, Australia's dragon fruit industry is relatively new, with potential for growth as interest in this exotic fruit

increases domestically. Dragon fruit thrives in regions with annual rainfall between 25 and 50 inches and tolerates temperatures up to 40 °C, making it suitable for cultivation in Australia's subtropical regions, particularly in Queensland and the Northern Territory (Hossain et al., 2021). Despite the growing global interest in dragon fruit as a functional food, research on its bioactive properties and therapeutic benefits, particularly within the context of Australian-grown varieties, remains sparse. This literature review seeks to address this gap by consolidating existing knowledge on dragon fruit's nutritional and bioactive composition, as well as its therapeutic potential and the status of global production standards. Unlike major global producers, the Australian dragon fruit industry operates without established quality benchmarks, limiting its ability to align with international frameworks and meet the increasing demand for health-promoting produce. This review is novel in its emphasis on the implications of these gaps for the Australian industry and the potential to integrate bioactive-focused standards.

By synthesizing existing findings, this work provides a foundation for the advancement of scientific understanding while supporting the development of industry standards that enhance the competitiveness and sustainability of Australian-grown dragon fruit. This comprehensive literature review explores classification and bioactive phytochemical composition of dragon fruit, emphasizing its nutritional and therapeutic properties. Moreover, it highlights the global dragon fruit production landscape and the current global industry standards for fresh produce, underscoring the need for Australia's emerging dragon fruit industry to align with international frameworks and capitalize on its competitive advantages by potentially establishing classifications for shape, size, defect tolerance, etc. This review aims to provide insights into the potential of establishing robust industry standards in Australia to enhance the marketability and sustainability of its dragon fruit production.

Published literature from the last two decades related to the cultivation, bioactive phytochemicals, nutritional value, therapeutic potential, and global industry standards for Hylocereus spp. (dragon fruit) was comprehensively reviewed. Relevant studies, including original research articles, review papers, and cross-referenced citations. were identified through numerous databases from the 'EBSCOhost' host platform, such as PubMed, SciFinder, Web of Knowledge, Scopus, and ScienceDirect. Search terms included combinations of (dragon fruit OR Hylocereus OR pitaya) AND (bioactive compounds, nutritional value, antioxidants, phenolic acids, flavonoids, betalains, anthocyanins, antimicrobial, anti-inflammatory, OR therapeutic properties). The inclusion criteria focused on articles that discussed the bioactive and therapeutic properties of dragon fruit and global production standards, with a specific emphasis on literature

evaluating quality benchmarks in Australia and other major producing regions. Global industry standards and Australian dragon fruit market reports were sourced directly from government sites and horticultural bodies such as AgriFutures Australia.

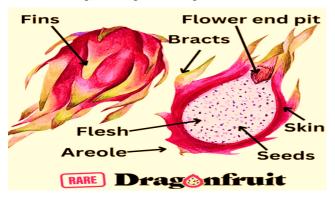


**Fig. 11:** Selected three species of dragon fruits (*Hylocereus* spp.) for analysis of nutrition and antioxidant.

Sources: DOI: 10.7717/peerj.17719/fig-1

## 5. Dragon Fruit Origin and Classification

Dragon fruit, also commonly referred to as pitaya (most commonly of the genus *Hylocereus*), belongs to the *Cactaceae* family, which comprises various cactus species. While this fruit is originally native to the arid regions of Mexico and northern South America, its commercial cultivation has spread widely beyond its native habitats. Nowadays, dragon fruit is extensively grown in tropical and subtropical regions around the world, including countries such as Vietnam, China, the Philippines, Israel, Australia, etc., where it has gradually become an important agricultural product (Chen *et al.*).



Sources: by Michael Schiffermuller, July 03, 2023

The plant can thrive under specific environmental conditions, with ideal growth occurring in areas that receive annual rainfall between 30 and 50 inches. It is also highly tolerant of elevated temperatures, capable of withstanding heat up to 40 °C. As a member of the cactus family, dragon fruit adapts well to arid to semi-arid

environments, making it particularly suited to regions with high light intensity and warm climates. However, while dragon fruit can survive in dry conditions, it requires a reliable supply of water to reach optimal growth. Additionally, the quality of the soil is a critical factor; the fruit flourishes best in fertile, well-drained soils that allow its root system to develop and support the plant's growth. Thus, despite its cactus-like resilience to heat and light, adequate moisture and soil fertility.

#### (i) Global Dragon Fruit Production

Major dragon fruit production regions are including China, Japan, Bangladesh, the Philippines, Vietnam, Australia, Indonesia, Sri Lanka, India, Thailand, South Africa, Spain, and the USA. Research on global dragon fruit production from 2017-2018 shows that Vietnam leads the world in dragon fruit production, with an impressive cultivation area of over 55,000 hectares and a total output of over 1,000,000 metric tons. Its productivity, ranging from 22 to 35 metric tons per hectare, highlights Vietnam's favorable conditions (Tarte et al., 2021). As a result, Vietnam dominates the global market in terms of both volume and productivity, capitalizing on its large-scale operations. China follows closely, with 40,000 hectares of land dedicated to dragon fruit farming, yielding 700,000 metric tons. China's productivity rate of 17.5 metric tons per hectare is slightly lower than Vietnam's but still reflects a strong level of production efficiency (Sharma et al., 2021). Indonesia, despite having a smaller production area of around 8500 hectares, impressively produces over 200,000 metric tons of dragon fruit, with a productivity rate of 23.6 metric tons per hectare (Sharma et al., 2021) In contrast, Thailand operates on a more modest scale, with a cultivation area of nearly 3500 hectares and a total production of 26,000 metric tons. Its productivity, at 7.5 metric tons per hectare, is one of the lowest among the major producers. Taiwan's dragon fruit industry is relatively small, with a cultivation area close to 2500 hectares and an annual production of 49,000 metric tons. Although Taiwan's production volume is lower than that of countries like Vietnam and Indonesia, its focus on maximizing yields (19.7 metric tons per hectare) from smaller areas, which ensures that its dragon fruit industry remains competitive. Malaysia and the Philippines have cultivation areas of 680 hectares and 485 hectares, respectively. Malaysia produces 7820 metric tons, with a productivity of 11.5 metric tons per hectare, while the Philippines yields 6062 metric tons, with productivity ranging between 10 and 15 metric tons per hectare and essential for the production of high-quality fruit. This adaptation to diverse climates, coupled with its ability to thrive in a range of soil conditions, has made dragon fruit a viable crop in many regions outside its native range, contributing to its rising global popularity as both an agricultural product and a health-promoting fruit.

 Table 8 : Status of Pitaya (Dragon Fruits).

Table 8: Status of Pitaya (Dragon Fruits).						
Origin	Color Classification	Varieties	Morphological Characteristics			
Israeli	red-skinned fruit with red flesh with greenish scales	Hylocereus costaricensis or Hylocereus polyrhizus (Pitaya roja or red-fleshed pitaya)	A medium to large berry with red peel and green or red fleshy scales.			
Colombia	yellow-skinned white flesh	Selenicereus megalanthus	The fruit is elongated ellipsoid with green scale ends. The yellow pericarp is particularly prominent.			
Antalya red-fleshed cultivars Türkiye		Dark Star [Hylocereus undatus], Philippines Purple [Hylocereus polyrhizus], Thai Red [Hylocereus polyrhizus], Malaysia Red [Hylocereus polyrhizus], Condor [Hylocereus guatemalensis], American Beauty [Hylocereus guatemalensis])	The fruit is typically round or oval in shape, uneven in size, and has a green, scaly appearance. Red skin and either red or white flesh are common.			
	white-fleshed cultivars	('Costa Rica White' [Hylocereus undatus] and 'Vietnamese Jaina' [Hylocereus undatus]).				
	red-fleshed, oval fruit	Selenicereus. monacanthus	The fruit comes in varying shapes, with most			
	red-fleshed,	Selenicereus sp.	being oval. It is available in three color combinations: red skin and red flesh, red skin			
Vietnam	white-fleshed, oval fruit	Selenicereus undatus	and white flesh, and yellow skin and white flesh. The fruit varies in size, has green scales			
	yellow-skinned, white-fleshed, oval fruit	Selenicereus megalanthus	at the tips, thicker skin, and dense seeds.			
	red-skinned with white flesh	Jumbo White and Vietnamese White	The fruit is round-ellipsoid in shape and			
Thai	white-skinned with white flesh	Pink, Siam Red, Taiwan Red, and Ruby Red	variably sized, with short scales evenly distributed. The rind of the pitaya is yellow			
	yellow or golden peel white flesh	Israel Yellow	and oval with green scale tips.			
	red peel with red pulp	Hongguan, Zhangjianghongrou, Guanhuahong, Hongshuijing	The fruit is oblong or ovoid, with red skin and either red or pink flesh, or white flesh. The			
	red-skinned with white pulp	Guanhuabai, Hongbaoshi, Yangxibairou	scales have sharply pointed green apices, and the fruit is 10–12 cm in length with thick, waxy skin. The umbilicus is small, and the flesh appears white, red, or pink in color. The			
China	red peel and pink pulp	Guanhuahongfen				
	red peel with bicolor pulp	Shuangse	scales have green tips, and the fruit size is uneven. This type of fruit is commonly			
	white-fleshed cultivars	('Costa Rica White' [Hylocereus undatus] and 'Vietnamese Jaina' [Hylocereus undatus]).	marketed as having red skin with white or red flesh.			
	red-skinned powder flesh	Hylocereus polyrhizus				
	powder-skinned white flesh	Hylocereus undatus	Common fruits found in the market include those that are oval or round in shape, have			
Bangladesh	yellow or golden peel white flesh	Israel Yellow	green scales, and have uneven sizes.  Additionally, some have red skin with pink			
	red flesh,	Selenicereus sp.	flesh, while others have pink skin with white			
	white flesh, oval fruit	Selenicereus undatus	flesh.			
	yellow skin, white flesh, oval fruit	Selenicereus megalanthus				
	red-skinned fruit with red flesh	Cebra (C, red, <i>Hylocereus. costaricensis</i> ); Lisa (L, red, <i>Hylocereus. costaricensis</i> ); Rosa (R, red, <i>Hylocereus. costaricensis</i> ); San Ignacio (SI, red, <i>Hylocereus. costaricensis</i> );	The fruit has a thicker skin and can be oval or round with green scales on the tips. There are three variations of the fruit: one with red skin and red flesh, with seeds uniformly and			
California	red-skinned	Physical Graffiti (PG, light pink, Hylocereus.	densely distributed; another with red skin,			
	powder flesh red-skinned, white flesh	polyrhizus and Hylocereus. undatus); Mexicana (M, white, Hylocereus. undatus)	pink flesh, and fewer, unevenly distributed seeds than the first variation; and a third with red skin, white flesh, and seeds densely distributed in the center of the fruit at the apex, with fewer seeds on the sides.			

Hylocereus spp. consists of several species, each displaying unique physical characteristics in terms of size, weight, and appearance. Hylocereus undatus (H. undatus) is one of the largest species in the group, with fruits ranging from 15 to 22 cm in length and weighing between 300 and 800 g. It is distinguished by its red peel contrasted with white flesh, giving it a visually appealing look. This species is widely recognized in the dragon fruit market due to its large size and striking coloration. Another notable species is Hylocereus polyrhizus (H. polyrhizus), which produces smaller fruits, typically measuring 10 to 12 cm in length and weighing 130 to 350 g. The peel of H. polyrhizus is red, and its flesh is also red, differentiating it from the white-fleshed *H. undatus*. The presence of green at the top of the fruit remains a shared feature within the Hylocereus genus, contributing to its overall aesthetic appeal (Le Bellec and Vaillant, 2011). Fruits ranging from 10 to 15 cm in length and weighing between 150 and 400 g. This species is unique for its light- or dark-red peel, with its red flesh creating a harmonious color scheme. Like other species in the genus, the green-tipped top adds visual consistency across the different types of Hylocereus fruit. Hylocereus costaricensis (H. costaricensis) is slightly larger and heavier, measuring 10 to 15 cm in length and weighing between 250 and 600 g. The peel is a vibrant red, while the flesh has a striking red-purple hue, making it distinct from the other red-fleshed varieties. The deeper color of the flesh further distinguishes *H. costaricensis* and adds to its visual and commercial appeal. Smaller species within the genus include Hylocereus trigonus (H. trigonus) and Hylocereus megalanthus (H. megalanthus), both of which produce fruits between 7 and 9 cm in length and weighing 120 to 250 g. Despite their similar sizes, these two species differ significantly in appearance. H. trigonus has white flesh, offering a contrast to the commonly red-fleshed species, while megalanthus is recognizable by its yellow peel and white flesh, a combination that sets it apart visually from the other species in the *Hylocereus* genus (Tarte *et al.*, 2021).

Cambodia and India, with relatively small cultivation areas, produce 4840 metric tons and 4200 annually, respectively. tons Cambodia's productivity stands at 11 metric tons per hectare, while India's ranges from 8.0 to 10.5 metric tons per hectare, suggesting that both countries could benefit from improved farming practices to boost their output. There is very limited literature on the Australian dragon fruit market, and it is still in its infancy, particularly when compared to larger global producers like Vietnam, China, and Indonesia. According to data from 2012–2015, Australia only produces 750 tons of fruit annually. As Australia's dragon fruit industry is relatively new, there is significant potential for domestic growth. With increased investment in research and the expansion of cultivation areas, Australia could enhance its productivity and meet rising local demand. The country's emphasis on sustainable farming practices and adherence to strict food safety standards gives it a competitive advantage, particularly in grown dragon fruit, this review focuses on global data to provide a general understanding. Most of the nutrients present in H. undatus and H. polyrhizus are recorded at higher levels than those in popular tropical fruits, including jack fruit, pineapple, and mango. It is worth noting that this Table 1 serves as a general comparison, as these values are highly influenced by factors such as geographical location, cultivation practices, and testing methodology (Liaotrakoon, W., 2013). The carbohydrate content in dragon fruit is similar to that of pineapple but lower when compared to other fruits, including banana, mango, and jack fruit. As a result, dragon fruit contains fewer calories, making it a suitable option for individuals aiming to lose weight (Arivalagan et al., 2021). Due to its relatively low sugar levels, dragon fruit is also considered a low-GI (glycemic index) fruit, beneficial for individuals with diabetes (Dayal et al., 2018). The main sugar present in dragon fruit is glucose (6 g/100 g), followed by fructose, and it also contains sorbitol, which contributes to its sweet flavour (Liaotrakoon, 2013) Additionally, dragon fruit is rich in vital minerals such as calcium, magnesium, potassium, iron, phosphorus, and sodium, all of which offer significant health benefits.

## (ii) Nutritional Value

#### **Chemical Composition**

Red-fleshed (H. polyrhizus) and white-fleshed (H. undatus) dragon fruits are the cultivars found the most in the Australian industry, and as such, only these two species are discussed in this section. These minerals play crucial roles in strengthening bones and teeth, regulating blood pressure and blood sugar, and aiding in the synthesis of essential amino acids (Ryan-Harshman and Aldoori, 2005). Dragon fruit is also a valuable source of vitamins, including thiamine (vitamin B1, 2.4 μg/100 g), riboflavin (vitamin B2, 2.0 μg/100 g), niacin (vitamin B3, 12.6 µg/100 g), and ascorbic acid (vitamin C, 5.6 µg/100 g) (Arivalagan et al., 2021). With a high moisture content of over 85%, the fruit's valuable source of vitamins, including thiamine (vitamin B1, 2.4 µg/100 g), riboflavin (vitamin B2, 2.0 µg/100 g), niacin (vitamin B3, 12.6 μg/100 g), and ascorbic acid (vitamin C, 5.6 μg/100 g) (Arivalagan et al., 2021). With a high moisture content of over 85%, the fruit's flesh is juicy and hydrating (Islam et al., 2012).

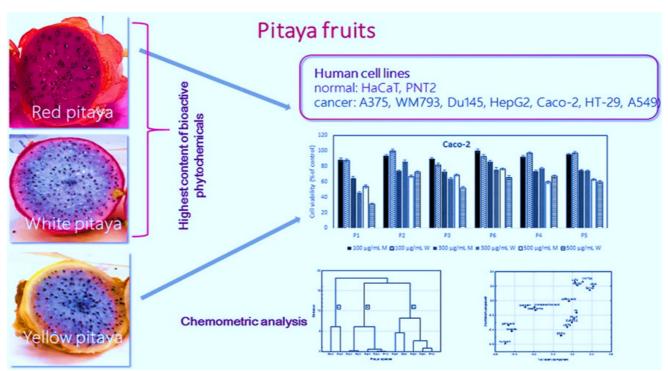
#### Bioactive Phytochemicals in Dragon Fruit

Phytochemicals, naturally occurring secondary metabolites found in various plant parts, are increasingly recognized for their significant bioactive properties and health benefits (Balamurugan, V.; Fatima, S.; Velurajan, S., 2019). Recent studies have shown that dragon fruit is

abundant in several important phytochemicals, including phenolic acids; flavonoids; and pigments such as carotenoids, betalains, anthocyanins, etc. These compounds are found in the flesh, seeds, and discarded peels of dragon fruit, which makes it a versatile source of bioactive components (Le and Huynh, 2021). As detailed in Table 2, *Hylocereus* species, including *H*.

Salicylic acid, although found only in the flesh, is primarily recognized for its antioxidant activity (Arivalagan *et al.*, 2021). Dragon fruit is also a rich source of hydroxycinnamic acids, which include p-coumaric acid, caffeic acid, chlorogenic acid, and sinapic acid. These compounds are highly valued for their antioxidant and anti-diabetic properties. P-coumaric acid, found in the flesh, has demonstrated both antioxidant and anti-diabetic activities (Tang *et al.*, 2021). Caffeic acid, present in the flesh and peel, is primarily known for its

antioxidant function (Dadhaneeya et al., 2021); Flavonoids are another key group of bioactive compounds in dragon fruit, including quercetin, catechin, rutin, phloridzin, and hesperidin. These flavonoids are abundant in the flesh, peel, and seeds and are recognized for their antioxidant, anti-diabetic, and anti-inflammatory properties. Quercetin, which is present in the flesh and peel of both H. polyrhizus and H. undatus, is known for its strong anti-inflammatory and antioxidant effects (Al-Mekhlafi et al., 2021; Castro-Enríquez et al., 2020; Le, 2022; Saenjum et al., 2021). Pires et al., 2022. Chlorogenic acid, located in the flesh, also contributes to anti-inflammatory and anti-diabetic effects (Arivalagan et al., 2021; Yan et al., 2020). Catechin, also a potent antioxidant, has been detected in the flesh and peel (Arivalagan et al., 2021; Fidrianny et al., 2017).



Rutin, which is present in the flesh and seeds, has been linked to anti-inflammatory, antioxidant, and anti-diabetic effects (Al-Mekhlafi *et al.*, 2021; Fidrianny *et al.*, 2017;2021). Phloridzin and hesperidin, found in both the flesh and peel, contribute additional antioxidant and anti-diabetic properties, with hesperidin also displaying anti-cancer potential (Ahmadi *et al.*, 2021; Chen *et al.*, 2021; Younis *et al.*, 2023). Carotenoids, particularly lycopene and  $\beta$ -carotene, are prominent in *H. polyrhizus*. Lycopene is known for its antioxidant and anti-cancer properties, while  $\beta$ -carotene offers antioxidant, anti-diabetic, and cardiovascular protective effects (Moo-Huchin, V.M.; Gonzlez-Aguilar, G.A.; Moo- antioxidant activity of these phytochemicals is determined by their ability to neutralize free radicals by donating electrons, thereby preventing cellular and tissue damage (Liaotrakoon, W. in the human body. These unstable molecules can cause like cancer and cardiovascular disorders.

## (iii) Antioxidant Capacities

The production of reactive oxygen species (ROS), commonly known as free radicals, is a key factor causing oxidative stress counteracting this damage by neutralizing free carotenoids, and betalains, which are found in the peel, flesh, and seeds. Research conducted by extensive damage to essential cellular components, such as proteins,

nucleic acids, lipids, and carbohydrates, potentially leading to mutations and the development of chronic diseases (Huchin *et al.*, 2021). Additionally, anthocyanins such as cyanidin-3-glucoside, delphinidin-3-glucoside, and pelargonidin-3-glucoside are vital components of dragon fruit, particularly in *H. polyrhizus* and *H. undatus*. These anthocyanins found in the flesh and peel exhibit

potent antioxidant and anti-inflammatory activities (Al-Mekhlafi *et al.*, 2021; Vargas *et al.*, 2013).

## (iv) Health Benefits of Dragon Fruit

For centuries, the medicinal properties of dragon fruit have been well recognized, with historical evidence revealing that the Mayan civilization used both its fruit and flowers for a variety of health-related purposes. These included managing blood sugar levels, promoting wound healing, acting as a diuretic, and treating conditions such as dysentery and tumors. Additionally, the seeds and flowers were often used to prepare beverages intended to alleviate gastritis, support bowel movements as a natural laxative, and enhance kidney function. The therapeutic benefits of dragon fruit are largely attributed to its rich concentrations of bioactive compounds predominantly found in the fruit itself.

Notably, dragon fruit has been proven to be non-toxic in scientific studies. Animal models administered doses as high as 5000 mg/kg of dragon fruit extract exhibited no harmful effects or organ abnormalities, underscoring its safety for potential medicinal applications (Kumar et al., 2018; Huang et al., 2021; Hernández and Salazar, 2012; Safira et al., 2021). This favorable safety profile, coupled with its extensive bioactive composition, makes dragon fruit a promising candidate for further exploration in the field of natural medicine. The subsequent sections delve into the various health benefits attributed to dragon fruit, including its antioxidant, anti-inflammatory, antimicrobial, anti-diabetic, anti-cancer, anti-lipidemic/antiobesity, anti-viral, and probiotic properties. These effects are supported by its bioactive compounds, which contribute to the fruit's diverse medicinal potential.

**Table 9:** Bioactive phytochemicals isolated from dragon fruit (*H. polyrhizus and H. undatus*) and their health benefits.

Phytochemical	Structure	Varieties	Parts	Health Benefits	
Hydroxybenzoic	Hydroxybenzoic acids				
Gallic acid	HO OH	H. polyrhizus and H. undatus	Flesh, peel, and seeds	Antioxidant, anti-obesity, and anti-diabetes effects	
Vanillic acid	HO CHAS	H. polyrhizus and H. undatus	Flesh and peel	Antioxidant, anti-diabetic, anti-atherogenic, and anti- inflammatory effects	
Syringic acid	HO OH	H. polyrhizus and H. undatus	Flesh, peel, and seeds	Antioxidant, anti-microbial, anti-cancer, anti- inflammatory, and anti- diabetes effects	
Salicylic acid	O O	H. polyrhizus and H. undatus	Flesh	Antioxidant effects	
Hydroxycinnami	c acids				
p-Coumaric acid	NO N	H. polyrhizus and H. undatus	Flesh and peel	Antioxidant and anti- diabetes effects	
Caffeic acid	HO	H. polyrhizus and H. undatus	Flesh and peel	Antioxidant effects	
Chlorogenic acid	HO OH	H. polyrhizus and H. undatus	Flesh	Antioxidant, anti- inflammatory, and anti- diabetes effects	

Sinapic acid	H <sub>3</sub> C OH	H. polyrhizus and H. undatus	Flesh and seeds	Antioxidant and anti- inflammatory effects
Flavonoids (Non-	-pigment)			
Quercetin	HO OH OH	H. polyrhizus and H. undatus	Flesh and peel	Antioxidant, anti-diabetes, and anti-inflammatory capacities
Catechin	HO OH OH	H. polyrhizus and H. undatus	Flesh and peel	Antioxidant effects
Rutin		H. polyrhizus and H. undatus	Flesh and seeds	Antioxidant, anti- inflammatory, and anti- diabetes effects
Phloridzin	HO HO HO HO	H. polyrhizus and H. undatus	Flesh and peel	Antioxidant, and anti- diabetes effects
Hesperidin		H. polyrhizus and H. undatus	Flesh	Antioxidant and anti-cancer effects
Carotenoids	H <sub>3</sub> CCH <sub>3</sub>		I	
Lycopene	CH <sub>3</sub>	H. polyrhizus	Flesh	Antioxidant, anti-cancer and anti-diabetes effects
β-carotene	CH <sub>3</sub>	H. polyrhizus	Flesh	Antioxidant, anti-diabetes, and anti-cardiovascular potential
Anthocyanin's	ÓН		<u> </u>	
Cyanidin-3- glucoside	HO OH OH	H. polyrhizus	Flesh and peel (including H. undatus)	Antioxidant and anti- inflammatory effects

Delphinidin-3- glucoside	HO OH OH	H. polyrhizus	Flesh and peel (including <i>H. undatus</i> )	Antioxidant and anti- inflammatory effects
Pelargonidin-3- glucoside	HO H	H. polyrhizus  Betalains	Flesh and peel (including <i>H. undatus</i> )	Antioxidant and anti- inflammatory effects
	0	Betalains	1	
Betacyanin	HO LO COLOR OF THE	H. polyrhizus	Flesh and peel	Antioxidant, anti-microbial, anti-viral, and anti- inflammatory effects
Betanin	HO H	H. polyrhizus	Flesh and peel	Antioxidant and anti- microbial capacities

**Note:** Concentration data are not included due to significant variability across studies influenced by environmental and agricultural factors and the lack of data specific to Australian-grown dragon fruit.

Charoensiri et al. identified beta-carotene and lycopene as major antioxidants in dragon fruit, both of which are effective in neutralizing free radicals (Charoensiri et al., 2009). Additionally, phenolic compounds, particularly flavonoids, are regarded as significant contributors to the fruit's overall antioxidant capacity and are highly concentrated in its edible parts (Recuenco et al., 2016). Manihuruk et al. demonstrated the antioxidant activity of H. polyrhizus peel, attributing it to the abundance of phenolic compounds, hydroquinones, and flavonoids (Manihuruk et al., 2017). Further evidence from Tenore et al. supports these findings, as they used 2,2-diphenyl-1-picrylhydrazyl (DPPH) and ferric-reducing antioxidant power (FRAP) assays to compare the antioxidant activity between dragon fruit peel and flesh. Their results indicated that the peel extract exhibited significantly higher antioxidant activity than the flesh due to a higher concentration of betacyanin in the peel) (Tenore, G.C.; Novellino, E.; Basile, A., 2012. Similarly, studies using DPPH assays found that the antioxidant properties of *H. undatus* and *H.* polyrhizus are dose-dependent, with the red-fleshed variety demonstrating greater antioxidant activity due to its higher levels of phenolic compounds and betacyanin (Mahayothee et al., 2019; Luo et al., 2014). Zulkifli et al. further reported that elevated processing temperatures can

enhance the antioxidant potential of the fruit (Yan et al., 2020).

Antioxidant properties in dragon fruit have been extensively studied across various plant parts, including the stem, leaves, flesh, peel, and seeds. The most abundant antioxidants in dragon fruit are phenolic compounds, betalains, and carotenoids, with alkaloids and vitamin C following in concentration (Lim *et al.*, 2007). This wide distribution of antioxidants contributes to the fruit's overall bioactivity, making dragon fruit a promising functional food for reducing the risk of chronic diseases such as cancer. In vivo studies have demonstrated that dragon fruit extract assists in neutralizing free radicals produced during exercise. This is reflected in decreased levels of oxidative stress markers such as creatine kinase in animal models (Harahap and Amelia, 2019).

The antioxidant capacity of dragon fruit is closely linked to its phenolic content, which is influenced by external factors such as solvent extraction and temperature. For instance, a comparative study of white-fleshed dragon fruit peel revealed that methanolic extraction yielded a significantly higher phenolic concentration than chloroform, providing a concentration of 48.2 mg GAE/100 g, as methanol can extract both polar and non-polar compounds. Chloroform, being

limited to non-polar compounds, resulted in lower antioxidant activity in DPPH assays (Som et al., 2019). Similarly, heat treatment ramping from 50 °C to 90 °C over one hour was shown to increase the antioxidant activities of dragon fruit, with almost double the total phenolic content determined in red-fleshed dragon fruit compared to the white-fleshed variety. Betacyanin, the primary antioxidant in red-fleshed dragon fruit, was identified as the main contributor to this enhanced activity, while the seeds also play an important role in maintaining antioxidant potential. Removal of the seeds significantly reduced the free radical-scavenging ability of the flesh (Liaotrakoon, 2013). Storage conditions also play a crucial role in preserving the antioxidant properties of dragon fruit. Research has shown that untreated dragon fruit experiences a slight increase in phenolic content during the first six days of storage, after which the content begins to decline. However, the addition of benzo (1,2,3)-thiadiazole-7-carbothioic acid S-methyl ester (BTH) to dragon fruit consistently exhibited increased phenolic and flavonoid contents, leading to enhanced antioxidant activity throughout storage (Ding et al., 2021). These results emphasize the significance of optimizing extraction methods and storage conditions to preserve dragon fruit's bioactive compounds maximize its therapeutic potential.

In short, dragon fruit is a rich source of natural antioxidants, including polyphenols, carotenoids, and betalains, which contribute to its potent free radical-scavenging abilities. The antioxidant potential of dragon fruit is influenced by various factors, such as the extraction method, environmental conditions, and storage practices. As research continues to explore the fruit's bioactive compounds, dragon fruit holds significant promise as a functional food capable of preventing oxidative stress-related diseases, supporting human health, and offering environmental resilience.

## (v) Anti-Inflammatory Capacities

Dragon fruit possesses notable anti-inflammatory properties, as shown in Figure 3. This figure summarizes the main anti-inflammatory effects of Hylocereus spp., which are discussed in subsequent sections. Antiinflammatory effects in dragon fruit are primarily due to the presence of bioactive phytochemicals such as anthocyanins, betalains, and squalene. Betalains, in particular, are recognized for their powerful antioxidant and anti-inflammatory effects. However, a significant challenge with betalains lies in their inherent instability under normal storage conditions. Environmental factors like oxygen, light, temperature, and pH greatly influence their stability, often leading to degradation and diminishing their effectiveness (Le et al., 2021). To overcome this issue, various strategies, such as encapsulating betalains in protective layers, have been proposed to enhance their stability and prolong their bioactive efficacy.

Rodriguez et al. derived betalains from the peel of red-fleshed dragon fruit and explored their antiinflammatory effects. Betalains were encapsulated in maltodextrin matrices, combining them with pectin and gum arabic to form a protective barrier to enhance their stability. The effectiveness of both encapsulated and nonencapsulated betalains in reducing inflammation was evaluated using a duck embryo chorioallantoic membrane (CAM) vascular irritation assay. In this assay, sodium dodecyl sulfate (SDS), a known irritant, was used to induce vascular irritation. The results showed that encapsulated betalains were five to six times more effective in reducing vascular irritation compared to nonencapsulated forms. This improved anti-inflammatory activity was attributed to the increased stability of encapsulated betalains, which retained their free radicalscavenging capabilities a key mechanism for reducing inflammation (Rodriguez et al., 2016).

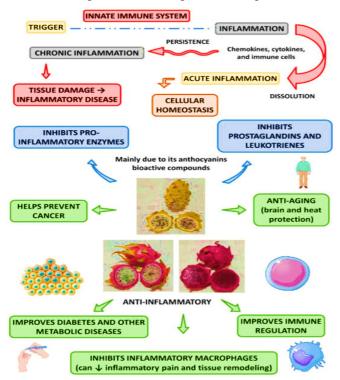
The anti-inflammatory action of betalains is closely tied to their antioxidant properties, as free radicals are major contributors to pro-inflammatory responses. By neutralizing these free radicals, betalains help to interrupt the inflammatory cascade, ultimately reducing tissue damage associated with chronic inflammation. Therefore, the ability of betalains to act as potent free radical scavengers plays a critical role in mitigating inflammation. Further research into the anti-inflammatory properties of the flesh and peel of dragon fruit, particularly white-fleshed H. undatus, has reinforced the bioactive potential of betalains. Eldeen et al. identified betalains as key contributors to the fruit's radicalscavenging and anti- Various studies have demonstrated the anti-microbial effects of dragon fruit, particularly redfleshed dragon fruit. One study assessed the ability of red dragon fruit to inhibit the growth of both Gram-negative and Gram-positive bacterial strains stored under cold conditions. The minimum inhibitory concentration (MIC) was measured by the broth microdilution method, with findings indicating that betacyanin levels in the fruit rose by almost 60% after 6 days of storage at 4 °C, followed by a gradual decrease. This increase was attributed to the hydrolysis of betalains into betalamic acid due to the fruit's water content. inflammatory capatities. Moreover, this research was the first to report the presence of squalene, a naturally occurring triterpene detected in the flesh of dragon fruit, with a concentration as high as Squalene exhibited considerable inflammatory activity by inhibiting important proinflammatory enzymes, including acetylcholinesterase, cyclooxygenase-2 (COX-2), and 5-lipoxygenase (5-LOX), with of EC50 values up to 47 µg/mL. Since these enzymes are involved in regulating inflammation and neural functions, their inhibition suggests that squalene may have therapeutic potential for th treatment of

inflammatory and neurodegenerative diseases (Eldeen et al., 2020).

The combined effects of betalains and squalene underscore the potential of dragon fruit as a natural anti-inflammatory agent, addressing both inflammation and related conditions, such as oxidative stress and neural disfunction. While betalains reduce inflammation by neutralizing free radicals, squalene strengthens this effect by inhibiting key pro-inflammatory enzymes. These findings indicate that dragon fruit, particularly when processed to preserve the stability of its bioactive compounds, holds promise as a therapeutic option for the management of inflammation, oxidative stress, and neural health.

#### (vi) Anti-Microbial Potential

The rise of antibiotic resistance and the growing prevalence of infectious diseases have sparked a pressing need for alternative anti-microbial agents. Conventional antibiotics are becoming less effective due to the development of microbial resistance, prompting a shift towards the exploration of natural anti-microbials from fruits and vegetables, which are less likely to induce side effects or contribute to drug resistance in microbes (Joshi, M.; Prabhakar, B. ,2020). Dragon fruit has shown great potential as a natural anti-microbial agent, displaying effectiveness against a wide range of microorganisms.



**Fig. 12:** Major anti-inflammatory effects of *Hylocereus* spp. (Nishikito *et al.*, 2023).

The study found that dragon fruit stored under cold conditions exhibited enhanced anti-microbial activity compared to freshly harvested fruit, which was linked to its higher betacyanin levels at the six-day mark (Yong *et al.*, 2018). These findings suggest that proper post-harvest storage conditions could improve the fruit's antimicrobial properties.

The anti-microbial properties of different parts of dragon fruit, including the peel, flesh, and whole fruit, have also been studied. When tested against 13 bacterial strains, two yeast strains, and four mold strains with the application of broth dilution, while whole fruit extracts were largely inactive, both the peel and flesh extracts exhibited significant anti-microbial activity. Polyphenolic compounds in the peel and flesh were identified as the primary contributors to this effect. Interestingly, Grampositive bacteria were more susceptible to the extracts than Gram-negative bacteria, likely due to the additional outer membrane in Gram-negative bacteria, which acts as a barrier against anti-microbial agents (Tenore *et al.*, 2012).

Recent innovations have further enhanced the antimicrobial capacities of dragon fruit, particularly through the use of silver nanoparticles synthesized from dragon fruit peel extracts. These nanoparticles were tested for antibacterial activity against pathogens such Escherichia coli, Staphylococcus aureus, Pseudomonas aeruginosa, with gentamicin serving as the positive control. A disc diffusion assay revealed that the silver nanoparticles exhibited significant antibacterial activity against all the tested pathogens. The stability and increased anti-microbial efficacy of these nanoparticles were credited to the phytochemicals found in the peel extract of dragon fruit (Phongtongpasuk et al., 2016). The use of silver nanoparticles represents a promising method for boosting the anti-microbial efficacy of natural extracts.

Another study explored the antibacterial effects of the leaf extract of H. undatus and H. polyrhizus against bacteria that cause meningitis, including Neisseria meningitidis, Listeria monocytogenes, and Streptococcus pneumoniae. Both extracts demonstrated dose-dependent inhibition of these bacterial strains, with leaf extract of H. polyrhizus showing stronger antibacterial activity than H. undatus. This increased efficacy was linked to the presence of flavonoids, which are known to disrupt bacterial cell walls by forming complexes, thereby inhibiting bacterial growth (Ritarwan and Nerdy, 2018). These results suggest that dragon fruit may have specific applications in combating bacterial infections, including meningitis. In short, these studies illustrate the broadspectrum anti-microbial potential of dragon fruit, driven primarily by its bioactive phytochemicals, such as betalains, polyphenols, and flavonoids. The peel and flesh have been shown to exert strong antibacterial effects against a variety of microbial strains, while advanced techniques such as nanoparticle synthesis have opened up new approaches to enhance the effectiveness of these

natural compounds. As antibiotic resistance continues to present a global health challenge, dragon fruit offers a promising natural alternative for the development of antimicrobial therapies that are both effective and less likely to contribute to resistance.

#### (vii) Anti-Diabetic Capacities

Characterized by persistent hyperglycemia, diabetes mellitus (DM) is a common metabolic disorder caused by either the pancreas failing to produce enough insulin or the body becoming less responsive to insulin. This condition leads to elevated blood glucose levels, which can severely impact a patient's overall health (Kaul et al., 2012). The dysfunction of insulin production or action is critical to the progression of DM. As the global incidence of diabetes continues to rise, the demand for effective treatment options has become urgent. Traditional medicines, particularly plant-based treatments, have attracted increasing attention due to their natural origins and reduced risks of side effects. Various medicinal plants have demonstrated promising results in managing diabetes by influencing key biochemical pathways, such as by inhibiting liver gluconeogenesis and improving cellular glucose uptake (Adinortey et al., 2019).

Among these natural therapies, the anti-diabetic properties of dragon fruit have attracted significant attention. Research indicates that both H. undatus and H. polyrhizus exhibit hypoglycemic effects that contribute to blood glucose regulation. For instance, Sudha et al. measured  $\alpha$ -amylase inhibition activity of H. undatus, which is a critical mechanism for controlling diabetes. The study revealed that as the amount of dragon fruit extract added to enzyme wells increased (from 25  $\mu$ L to 100  $\mu$ L),  $\alpha$ -amylase inhibition activity rose by 1% to 30%, highlighting the significant anti-diabetic potential of the fruit (Sudha et al., 2017).

In addition to white-fleshed dragon fruit, red-fleshed varieties have also been studied for their anti-diabetic effects. Abd Hadi et al. conducted a study on type 2 diabetic patients, showing that consuming 400 g of red dragon fruit daily significantly lowered triglyceride levels, while consuming 600 g per day helped reduce hyperglycemia, decrease total and LDL (low-density lipoprotein) cholesterol levels, and increase HDL (highdensity lipoprotein) cholesterol levels. These results emphasize red dragon fruit's potential to regulate blood glucose in individuals with type 2 diabetes (Abd Hadi et al., 2016). Further research into dragon fruit's antidiabetic properties has expanded understanding of its role in metabolic regulation. H. polyrhizus has been extensively studied for its effects on insulin resistance. Omidizadeh et al. investigated the impact of H. polyrhizus on insulin-resistant rats. This study involved feeding the rats both fresh and heat-processed dragon fruit samples with varying levels of phenolic contents and antioxidant activity. After six weeks, the rats exhibited a

significant reduction in insulin resistance, alongside improved atherosclerotic changes and lower hypertriglyceridemia. The polyphenols, soluble dietary fiber and antioxidant compounds in dragon fruit were credited for these effects, suggesting that the fruit could be beneficial in reducing insulin resistance through dietary intervention (Omidizadeh *et al.*, 2014).

Clinical trials on diabetic patients have further explored the effects of dragon fruit on lipid metabolism and glycemic control. A 7-week clinical trial conducted by Joshi *et al.* investigated the effects of *H. polyrhizus* consumption on blood glucose levels and lipid profiles in patients with type 2 diabetes. The study consisted of three phases pre-treatment, treatment, and post-treatment phases and found that consuming 400 g of dragon fruit daily significantly lowered triglyceride levels, while 600 g per day had a stronger effect in terms of reducing blood glucose levels. These results indicate that dragon fruit can positively influence both lipid metabolism and glycemic control in type 2 diabetes patients (Abd Hadi *et al.*, 2016).

A study by Poolsup et al. also emphasized the hypoglycemic impacts of both H. undatus and H. polyrhizus in individuals with diabetes and pre-diabetes. The analysis revealed that dragon fruit was more effective at reducing blood glucose levels in pre-diabetic individuals, with greater reductions observed at higher consumption levels (Poolsup et al., 2017). This dosedependent trend suggests that increasing dragon fruit intake may be beneficial in managing early stages of diabetes or preventing the development of type 2 diabetes. The in vitro hypoglycemic activity of methanolic extracts of dragon fruit was also demonstrated by Ravichandran et al. using protein glycation inhibition assays. The polyphenolic content of dragon fruit was found to be a key factor in its anti-glycation and antioxidant effects (Ravichandran et al., 2021). Although increasing evidence supports the use of dragon fruit in the management of diabetes and its related complications, further research, particularly human clinical trials, is needed to confirm its efficacy. Most studies have focused on animal models or in vitro experiments, providing valuable insights, but more clinical validation is necessary to determine dragon fruit's therapeutic potential for diabetic patients. Future research should focus on establishing optimal consumption patterns, dosages, and long-term effects to provide clearer guidance on incorporating dragon fruit into diabetes management strategies. In conclusion, dragon fruit shows considerable promise as a natural anti-diabetic agent. Dragon fruit's potential to enhance insulin sensitivity, regulate blood glucose levels, and manage lipid profiles positions it as a promising candidate for diabetes treatment. However, additional human trials are necessary to thoroughly understand its mechanisms and confirm its therapeutic benefits in clinical applications.

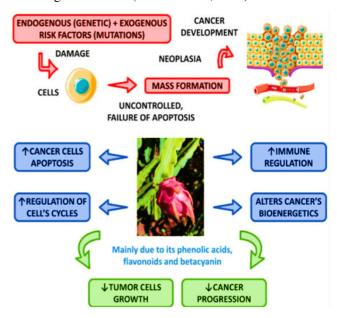
#### (viii) Anti-Cancer Potential

Imbalance between reactive oxygen species (ROS) and antioxidants in the body causes oxidative stress, a major contributor to the development of degenerative conditions such as cancer and Alzheimer's disease. This stress damages cellular structures, promoting mutagenesis and uncontrolled cell growth. Rich in bioactive compounds such as polyphenols, betalains, tocopherols, and unsaturated fatty acids, dragon fruit has exhibited chemotherapeutic potential against various cancer cell lines (Joshi and Prabhakar, 2020; Ibrahim *et al.*, 2018). These bioactive components work synergistically to reduce oxidative stress and inhibit cancer cell proliferation.

Dragon fruit's strong antiproliferative properties are largely attributed to its phenolic compounds. These compounds exhibit significant anti-cancer effects by targeting several biochemical pathways. Figure 4 shows the anti-cancer potential associated with *Hylocereus* spp. For instance, betalains, which are abundant in dragon fruit, have been found to inhibit lipid peroxidation and suppress cyclooxygenase (COX-1 and COX-2) enzymes, key players in inflammation and cancer progression. By disrupting these pathways, betalains help prevent the growth of tumor cells (Naseer, S.; Hussain, S.; Abid, A., 2019). Le *et al.* highlighted the role of phenolic compounds in enhancing dragon fruit's antiproliferative activity, suggesting that these compounds are critical to its anti-cancer effects (Le *et al.*, 2021).

Various studies have evaluated both cytotoxic and antiproliferative effects of dragon fruit against several cancer cell lines. For example, strong anti-cancer effects on colon and prostate cancer cells have been observed in both methanolic and aqueous extracts of dragon fruit, suggesting its potential as a natural therapeutic agent (Paśko et al., 2021). Additionally, research on whitefleshed dragon fruit (H. undatus) revealed that with increases in concentrations of dragon fruit extract, a significant decrease was observed in the viability of human hepatocellular carcinoma (HEPG-2) cells, underscoring the dose-dependent nature of its cytotoxic effects on liver cancer cells (Padmavathy et al., 2021). Another study conducted using a 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium (MTS) assay confirmed the strong cytotoxic effects of dichloromethane peel extracts, further highlighting dragon fruit's potential in cancer treatment (Hendra et al., 2021). A comparative study examined the effects of H. polyrhizus and H. undatus peel extracts on human breast, prostate, and gastric cancer cell lines, revealing dose-dependent cytotoxicity. The results showed that *H. polyrhizus* exhibited stronger inhibition, especially against human gastric cancer (MGC-803) cells, with inhibition rates exceeding 60% across all tested cell lines (Luo et al., 2014). The superior anti-cancer activity

of *H. polyrhizus* was attributed to its higher concentrations of pentacyclic triterpenoids and steroids, which are believed to contribute to its greater efficacy in inhibiting cancer cells (Ibrahim *et al.*, 2018).



**Fig. 13 :** Major anti-cancer effects of *Hylocereus* spp. (Nishikito *et al.*, 2023).

In addition to these compounds, other phytochemicals in dragon fruit, such as  $\alpha$ -amyrin and  $\beta$ amyrin, also exhibit anti-cancer and antiproliferative effects. However, α-amyrin was found to have relatively weaker effects compared to the other compounds when tested against breast, prostate, and gastric cancer cell lines (Luo et al., 2014). Kim et al. further supported these findings, ranking the anti-cancer efficacy of different dragon fruit extracts, with white-fleshed dragon fruit peel showing the highest activity, followed by red-fleshed peel and red-fleshed dragon fruit flesh. The peel of H. undatus was found to possess chemotherapeutic compounds, including flavonoids and polyphenols, at concentrations three to five times greater than other sources, which contributed to its superior antiproliferative activity.

Nitric oxide (NO), a well-known factor in promoting tumor growth through angiogenesis, also contributes to the development of cancer. Jayakumar  $et\ al.$  demonstrated that the proliferation of MCF-7 breast cancer cells increased by more than 150% following treatment with sodium nitroprusside, a nitric oxide donor. However, pretreatment with  $H.\ undatus$  extract significantly reduced cell viability, with a 600 µg/mL dose inhibiting MCF-7 cell growth by over 80%. The study attributed this effect to the phenolic compounds in dragon fruit, which exhibited strong free radical-scavenging activity, suppressing tumor growth (Jayakumar and Kanthimathi, 2011).

Dragon fruit has also been shown to mitigate the toxic side effects of conventional cancer treatments. In one study, Wistar rats treated with dragon fruit juice concentrate prior to cisplatin chemotherapy experienced protection significant against cisplatin-induced nephrotoxicity. This protective effect was linked to the high betanin content in the juice, which preserved renal function and reduced lipid peroxidation levels (Ramírez-Rodríguez et al., 2022). In recent years, dragon fruit has also been explored as a component in nanotechnologybased cancer therapies. Divakaran et al. synthesized dragon fruit extract-capped gold nanoparticles (DF-AuNPs) using a green method. These nanoparticles exhibited high levels of cytotoxic activity against MCF-7 breast cancer cells. The nanoparticles, measuring between 10 and 20 nm, selectively targeted MCF-7 cells, demonstrating dragon fruit extract's potential as both a capping and reducing agent in nanoparticle synthesis. This innovative approach may open new avenues for more targeted and effective cancer treatments (Divakaran et al., 2019).

In summary, extensive research highlights dragon fruit's antiproliferative and cytotoxic potential against various cancer cell lines. Its bioactive compounds such as polyphenols, betalains, flavonoids, and triterpenoids are especially abundant in the peel and contribute to its chemotherapeutic properties. Dragon fruit's ability to suppress tumor growth, reduce chemotherapy side effects, and facilitate novel therapies like nanoparticle synthesis underscores its multifaceted role in cancer treatment. However, while promising, further in vivo studies and clinical trials are needed to fully validate dragon fruit's therapeutic potential and ensure its safe and effective use in cancer treatment.

## (ix) Anti-Obesity Capacities

Dyslipidemia and obesity are two major risk factors for the development of cardiovascular diseases, as they both contribute to the onset of atherosclerosis and other vascular complications (Tsai *et al.*, 2019). Given that cardiovascular diseases remain one of the leading causes of death worldwide, there has been increasing interest in using natural products as therapeutic agents to manage these conditions. Various fruits and plant-based products have been extensively studied for their potential to regulate lipid metabolism and assist with weight management.

Dragon fruit (*H. polyrhizus*) has shown great promise in reducing cardiovascular risks. A study conducted by Hernawati *et al.* examined the effects of the peel powder of *H. polyrhizus* on lipid profiles, including total cholesterol, HDL, LDL, and triglycerides, in hyperlipidemic male mice. The study followed a randomized design, where mice were fed a high-fat diet for 20 days to induce hyperlipidemia, followed by treatment with different doses of dragon fruit peel powder

(50, 100, 150, and 200 mg/kg BW/day) for 30 days. Blood samples were collected to monitor lipid levels, and the results showed significant decreases in total cholesterol, LDL, and triglyceride levels, along with increases in HDL levels, particularly in the groups receiving higher doses. The peel's high crude fiber content (nearly 70%), which includes both soluble and insoluble fibers, was believed to contribute to these lipidlowering effects. Soluble fibers bind to bile acids and cholesterol, enhancing their excretion, while insoluble fibers reduce fatty acid synthesis in the liver. Additionally, dragon fruit peel contains tocotrienols and antioxidants, which help regulate lipid metabolism and improve insulin sensitivity (Setiawan et al., 2013). Another study explored the anti-lipase activity of H. undatus juice. Lipase, an enzyme involved in fat digestion, was inhibited by dragon fruit juice in a concentration-dependent model, with inhibition rates ranging from 6% to 46%, as assessed by the Rhodamine agar plate method (Sudha et al., 2017). By inhibiting lipase, dragon fruit juice may reduce fat absorption, offering a natural means of managing obesity and promoting weight loss.

In addition to these lipid-lowering effects, red dragon fruit has been found to counteract dyslipidemia, hypertriglyceridemia, and atherosclerosis in animal models. A study conducted by Omidizadeh *et al.* showed that consuming red dragon fruit reduced atherosclerotic changes in rats fed a fructose-rich diet. This was attributed to the high levels of soluble dietary fibers, polyphenols, and antioxidants present in the fruit, which collectively improved lipid metabolism and reduced oxidative stress (Omidizadeh *et al.*, 2014). Polyphenols, in particular, act as antioxidants that neutralize free radicals and prevent lipid peroxidation, protecting against vascular damage and reducing cardiovascular risk.

Dragon fruit's beneficial effects on lipid metabolism are closely linked to its nutrient-rich composition. The fruit is abundant in dietary fibers, antioxidants like tocotrienols, and bioactive polyphenols, all of which support healthier cholesterol levels and prevent fat accumulation. Furthermore, dragon fruit's ability to improve insulin sensitivity may help mitigate the risk of type 2 diabetes, which is often associated with dyslipidemia and obesity. Its inhibitory effect on  $\beta$ -Hydroxy  $\beta$ -methylglutaryl-CoA (HMG-CoA) reductase, an enzyme critical for cholesterol synthesis, and its ability to bind bile acids, promoting cholesterol excretion, further underscore its role as a functional food for the management of metabolic health.

While these animal studies present promising results, further research in human clinical trials is necessary to confirm dragon fruit's long-term effects on lipid metabolism and cardiovascular health. Future studies should establish proper consumption guidelines

and explore the therapeutic benefits of dragon fruit in preventing obesity and dyslipidemia. Despite the need for more evidence in humans, current research points to dragon fruit's considerable potential as a natural agent for improving metabolic health and preventing cardiovascular diseases.

Overall, dragon fruit offers significant promise in managing obesity and dyslipidemia due to its rich contents of dietary fibers, antioxidants, and polyphenols. Its ability to lower total cholesterol, LDL, triglycerides, and body weight while raising HDL levels highlights its multifaceted role in promoting cardiovascular health. Additionally, its capacity to inhibit lipase activity and improve insulin sensitivity positions dragon fruit as a promising natural intervention for the management of obesity and related metabolic disorders. Further studies should focus on validating these effects in human populations and determining optimal doses for therapeutic benefits.

#### (x) Anti-Viral Potential

Dragon fruit, particularly red-fleshed species, has drawn considerable attention for its anti-viral potential, primarily due to the presence of betacyanins, which are potent bioactive compounds. Several studies have explored the anti-viral potential of red dragon fruit, with promising results against various viral pathogens. Chang et al. conducted a key study that explored the anti-viral properties of betacyanins derived from H. polyrhizus in relation to the type 2 dengue virus (DENV-2), which is responsible for severe dengue fever cases. Dengue, which is a mosquito-borne illness, poses a serious global health threat, with DENV-2 being one of the most virulent strains. The study employed a plaque reduction neutralization test to evaluate how betacyanins could inhibit viral replication. Using a viral strain isolated from infected patient, the researchers ranked the concentrations of betacyanins in red dragon fruit as follows: phyllocactin (highest), betanin, isobetanin, and hylocerenin (lowest). This study also revealed that the anti-viral effect was dose-dependent, showing an IC<sub>50</sub> (half-maximal inhibitory concentration) of 125.8 µg/mL and a selectivity index (SI) of 5.3 µg/mL. These results highlight betacyanins' potential as natural anti-viral agents, particularly against DENV-2 (Chang et al., 2020). The study provided compelling evidence of the efficacy of red dragon fruit in reducing viral yield, suggesting its usefulness in combating dengue virus infections.

Red dragon fruit has also been explored for its antiviral capacities against SARS-CoV-2 (severe acute respiratory syndrome coronavirus 2), the virus responsible for the COVID-19 (Coronavirus disease 2019) pandemic, in addition to its potential efficacy against dengue. A molecular docking study examined the interaction between betacyanins and critical SARS-CoV-2 targets, including the receptor-binding domain (RBD)

of the spike protein and the major protease (Mpro). The demonstrated strong interactions betacyanins and these viral receptors, suggesting that these compounds could inhibit viral entry and replication. Various molecular interactions, such as hydrogen bonds and van der Waals forces, further supported the potential of betacyanins as therapeutic agents for SARS-CoV-2. These findings point to the possibility of using betacyanin-rich dragon fruit extracts as natural inhibitors of coronaviruses, opening new research approaches for the treatment and prevention of viral infections such as COVID-19 (Tallei et al., 2021). Moreover, betacyanins from red dragon fruit have shown anti-viral potential against other respiratory viruses, such as influenza. A study found that treatment with betacyanins significantly reduced virus titers in cells infected with influenza A, which is a common cause of seasonal flu and pandemics. This suggests that betacyanins could help lower viral loads and limit the spread of influenza within the host (Lim et al., 2024).

The body of research on dragon fruit underscores its strong anti-viral potential, particularly through the bioactive effects of betacyanins. These compounds appear to inhibit viral replication by interacting with viral receptors and possibly modulating the host's immune response to infection. However, while these studies present promising in vitro and molecular docking results, additional research is necessary to gain a comprehensive understanding of the mechanisms of the anti-viral effects betacyanins. Moving forward, studies should concentrate on in vivo models and clinical trials to evaluate the safety, effectiveness, and pharmacokinetics of betacyanins in the treatment of viral infections. Additionally, exploring the synergistic effects of betacyanins with other anti-viral agents could pave the way for combination therapies, maximizing the therapeutic potential of dragon fruit in combating viral diseases.

#### **Prebiotic and Probiotic Properties**

The health of the gut microbiota is essential for maintaining digestive health, and imbalances in this microbial community can lead to various gastrointestinal issues, such as diarrhea, constipation, inflammatory bowel disease (IBD), and irritable bowel syndrome (IBS). Prebiotics and probiotics play a significant role in promoting gut health, with prebiotics serving as nourishment for beneficial bacteria and probiotics contributing by replenishing these microbes (Chlebicz-Wójcik and Śliżewska, 2021). Dragon fruit, known for its rich prebiotic fiber content, has emerged as a promising natural remedy for supporting gut health and managing digestive disorders.

The primary carbohydrates in both *H. polyrhizus* and *H. undatus* are glucose, fructose, and oligosaccharides, with the latter being the most critical to dragon fruit's

prebiotic properties. These oligosaccharides resist hydrolysis by human α-amylase and artificial gastric juices, allowing them to pass through to the colon intact. In the colon, they stimulate the growth of beneficial bacteria, including Lactobacillus and Bifidobacterium, demonstrating clear prebiotic activity (Wichienchot et al., 2010). This ability to support beneficial bacteria suggests that dragon fruit can maintain a healthy gut microbiome by promoting the proliferation of these helpful microorganisms. Several studies have explored the prebiotic effects of oligosaccharides in dragon fruit. For example, Dasaesamoh et al. found that the fecal fermentation of dragon fruit oligosaccharides increased the populations of Bifidobacterium and Lactobacillus while reducing harmful bacteria such as Bacteroides and Clostridium. The fermentation process also produced short-chain fatty acids (SCFAs) like lactic acid, acetic acid, propionic acid, and butyric acid, which play vital roles in gut health. These SCFAs provide energy to colonocytes and contribute to anti-inflammatory processes, improved gut barrier function, and better metabolic health (Dasaesamoh et al., 2016). The production of SCFAs underscores the significant prebiotic potential of dragon fruit in supporting gut health and regulating digestive function.

Animal studies have further explored dragon fruit's prebiotic properties. Khuituan *et al.* conducted research on how oligosaccharides extracted from dragon fruit influence male ICR/Mlac mice. Results showed that these oligosaccharides increased transit time in the upper gastrointestinal tract while reducing total gut transit time. Additionally, they acted as both bulk-forming and stimulant laxatives by increasing fecal mass and enhancing gut motility, which is particularly beneficial for the management of conditions like constipation. These results exhibited that dragon fruit oligosaccharides offer both prebiotic benefits and therapeutic effects for gastrointestinal conditions (Ibrahim *et al.*, 2018; 2022).

The prebiotic properties and related compounds of both H. polyrhizus and H. undatus have been quantified using high-performance liquid chromatography (HPLC), with red dragon fruit containing slightly more oligosaccharides (90 g/kg) than white dragon fruit (86 g/kg). These oligosaccharides serve as a carbon source for beneficial study, Lactobacillus bacteria. In one delbrueckii experienced significant growth cultivated with dragon fruit oligosaccharides, increasing from  $9.0 \times 10^7$  to  $6.2 \times 10^9$  cells/mL in less than 48 h. Bifidobacterium bifidum also showed considerable growth, further confirming dragon fruit's effectiveness as a prebiotic. These results highlight the role of dragon fruit in promoting a balanced gut microbiome and supporting digestive health. Khalili et al. further confirmed the prebiotic properties of dragon fruit, revealing that oligosaccharides were more concentrated in the flesh of both H. polyrhizus and H. undatus than in the peel. They

noted that red dragon fruit exhibited slightly greater prebiotic effects than white dragon fruit. This emphasizes the importance of consuming the fruit's flesh to fully benefit from its prebiotic properties (Naseer *et al.*, 2019).

The positive impact of dragon fruit on gut health has broader implications beyond digestion. A balanced gut microbiota plays a crucial role in supporting immune function, reducing systemic inflammation, and regulating metabolic processes. Dragon fruit's ability to increase SCFA production especially the production of butyric acid, which is linked to reduced inflammation and improved insulin sensitivity suggests that it could help manage metabolic disorders like obesity and type 2 diabetes. Additionally, dragon fruit's role in promoting a diverse and balanced microbiome may contribute to the prevention of gut dysbiosis, a condition associated with various diseases, including autoimmune disorders and mental health conditions.

#### 6. Uses of Processed Dragon Fruit

The importance of dragon fruit has been recognized not only for its nutritional value but also in folklore and traditional medicine. The fruit can be consumed in its raw form or processed into a variety of products, including juice, wine, spreads, and desserts. In traditional herbal medicine, dragon fruit has been valued for its therapeutic properties, especially in promoting digestive health and combating various ailments. Despite its multifaceted benefits, dragon fruit remained largely underappreciated and undervalued for many years. However, recent scientific investigations have shifted this perspective, unveiling a wealth of evidence supporting the potential health benefits of dragon fruit. Research has highlighted its significance in addressing inflammation, cancer, and diabetes, alongside its potential as a natural colorant in food and cosmetics (Lwin et al., 2020). Furthermore, the peels of the fruit have been found to possess a strong capacity to absorb toxins, further contributing to its value in food and environmental safety (Nurmahani et al., 2012).

## (i) Food and Cosmetic Industry

One of the most remarkable industrial uses of dragon fruit lies in its high betalain content. Betalains are natural, water-soluble pigments found abundantly in the fruit, particularly in the *H. polyrhizus* species. These pigments are valued for their ability to add vibrant colors to food products without affecting taste. In addition to their coloring properties, betalains are rich in proteins, fats, fibers, and antioxidants, making them a valuable component in both the food and health industries (Tze *et al.*, 2012).

Dragon fruit peel has shown considerable potential as a natural thickening agent in the food industry. Studies demonstrated that at 5% concentrations, alcohol air residues derived from dragon fruit peels exhibited

viscosity properties comparable to those found in commercial thickening agents, suggesting that dragon fruit peel could serve as an effective alternative in food product formulations (Montoya-Arroyo *et al.*, 2014). Additionally, the cosmetics industry has increasingly adopted betalains from dragon fruit as a natural alternative to synthetic colorants and chemicals, which are often associated with skin irritations and allergic reactions. Betalains provide various skin benefits without the risk of adverse side effects, making them a popular choice in skincare formulations (Kamairudin *et al.*, 2014).

Another valuable component of dragon fruit is its seeds, which are rich in essential oils. Both H. polyrhizus and H. undatus seeds contain significant amounts of oil, ranging from 18.3% to 28.4%, with notable total tocopherol contents of 36.7 mg/100 g and 43.5 mg/100 g, respectively (Lim et al., 2010). Research has demonstrated that dragon fruit seed oil is particularly high in linoleic acid, which is an omega-6 fatty acid, surpassing the concentrations found in other seed oils like canola, linseed, sesame, and grapevine oils. The oil from white-fleshed and red-fleshed dragon fruit seeds contains 50% essential fatty acids, with linoleic acid contributing 48.5% and linolenic acid contributing 1.5% of the oil composition (Ariffin et al., 2009). Another study reported linoleic acid contents of 540 g/kg for H. undatus and 480 g/kg for H. polyrhizus, confirming the oil's richness in functional lipids. These findings highlight the potential of dragon fruit seed oil as a valuable source of essential oils, with applications in both nutritional and cosmetic products.

#### (ii) Others

Dragon fruit's versatility extends across multiple industries, offering valuable applications in food, beverage, and dairy production. For example, fruit wine made from *H. polyrhizus* contains a rich array of aroma compounds, such as esters (66.2%), alcohols (18.2%), alkanes (4.3%), acids (5.9%), aldehydes (0.1%), olefins (0.1%), and other volatile substances (0.2%) (Gong *et al.*, 2017). This diverse composition of volatile compounds contributes to the wine's unique flavor profile, making it an attractive product for the beverage industry.

In the dairy sector, the addition of flesh of whitefleshed and red-fleshed dragon fruit into yogurt has been shown to improve its nutritional qualities. Adding dragon fruit flesh enhances lactic acid and total phenol contents, as well as antioxidant activity, while also accelerating the milk fermentation process (Zainoldin, 2012). These enhancements not only boost the nutritional value of yogurt but also contribute to its classification as a functional food, offering additional health benefits.

In the baked goods industry, dragon fruit peel flour has been explored as a partial replacement for wheat flour in cookie recipes. When 15% dragon fruit peel flour was

added to wheat flour, the resulting cookies showed increased ash and fiber contents, as well as higher carbohydrate levels, larger diameters, and a greater spread ratio (Ho and Abdul Latif, 2016). This suggests that dragon fruit peel flour can improve the nutritional profile of baked goods, making it a valuable ingredient for the creation of healthier snack options.

Furthermore, red-fleshed dragon fruit peel powder has been proposed as a fat substitute in ice cream formulations, particularly for those seeking low-calorie alternatives. By reducing the fat content and adding dietary fiber, dragon fruit peel powder enhances the nutritional value of ice cream while lowering its caloric content (Nizamlioğlu *et al.*, 2012). This innovative use of dragon fruit peel demonstrates its potential in producing healthier, calorie-reduced food products.

## 7. Dragon Fruit Industry

## (i) Overview

The Australian dragon fruit market is relatively small but steadily growing, with most production concentrated in Queensland, New South Wales, South Australia, and the Northern Territory. Dragon fruit, or Hylocereus spp., was introduced to Queensland in the 1970s, and cultivation has since spread to other regions of the country. The primary varieties grown in Australia are white-fleshed H. undatus and red-fleshed H. polyrhizus. However, despite being an economically important fruit with multiple uses, dragon fruit cultivation in Australia remains niche. It accounts for a relatively small portion of the nation's horticultural output, yielding around 740 tons annually from approximately 40,000 plants (Chen et al., 2024). The market for locally grown dragon fruit is further limited by high production costs, which make it difficult for Australian growers to compete on price with cheaper imported dragon fruit from Southeast Asia. Despite the growing interest in dragon fruit, there are still limited amounts of literature and data available specifically focused on the Australian dragon fruit industry. Much of the current research on dragon fruit centers on larger global producers such as Vietnam, Thailand, and other Southeast Asian countries. In contrast, the Australian industry remains underresearched, and there is a distinct lack of comprehensive studies detailing the market dynamics, consumer preferences, and trade patterns specific to Australia. This lack of detailed information poses challenges for local growers, who need accurate data to make informed decisions about production, marketing, and strategic planning. Without sufficient research and robust industry data, the Australian dragon fruit industry is left at a disadvantage when attempting to compete in both domestic and global markets (Abhishek Soni, 2014).

#### (ii) Imported Dragon Fruit

A significant challenge faced by the Australian dragon fruit market is the increasing competition from imports, particularly from Vietnam. Since 2017, the influx of imported dragon fruit has led to a noticeable drop in the price of Australian-grown produce. Figure S1 in the Supplementary Materials compares the monthly average prices of Australian-grown dragon fruit with those of imported dragon fruit over a six-year period from 2014 to 2020. The figure shows that the price of Australian-grown dragon fruit remained consistently higher than that of imported dragon fruit throughout the period. This price difference reflects the higher production costs in Australia, particularly labor and resource expenses, compared to lower-cost countries such as Vietnam. The monthly fluctuations in prices also reveal seasonal trends. The prices of Australian-grown dragon fruit typically peak during the off-peak seasons, when local supply is lower and imported fruit is more abundant. Figure S2 in the Supplementary Materials illustrates a critical turning point in the Australian dragon fruit market, showing the monthly average prices of Australian-grown and imported dragon fruit both before and after 2017, which is when imports from Southeast Asia, particularly Vietnam, were first introduced, providing a clear comparison of trends over time (Abhishek Soni, 2014). The figure highlights a distinct drop in the price of Australian-grown dragon fruit following the commencement of imports in 2017. Before 2017, the prices of Australian-grown dragon fruit were relatively stable, with occasional seasonal fluctuations. However, after 2017, there was a notable downward trend in prices, reflecting the impact of cheaper imported fruit on the local market. The figure underscores the significant market disruption caused by the introduction of imports, which placed additional pricing pressure on local producers and contributed to the ongoing challenges faced by the Australian dragon fruit industry.

#### (iii) Current Issues

One of the most pressing issues in the Australian dragon fruit industry is the relatively high production costs compared to imported fruit. Labor and resource expenses are significantly higher in Australia, making it difficult for local farmers to produce dragon fruit at competitive prices (Weller, S.A.). Additionally, the introduction of cheaper imported fruit has created downward pressure on prices, which continues to impact the profitability of Australian-grown dragon fruit. Australian growers also face limited opportunities for export due to strict biosecurity regulations and high costs associated with exporting to international markets (Richards and Higgins, 2016).

Moreover, there is a lack of research into off-peak production methods, such as controlled environment agriculture, which could help increase the availability of Australian dragon fruit throughout the year. Currently, Australian growers are limited to peak production seasons, making it harder to compete with year-round imported fruit. Technologies including light stimulation for flowering and temperature control during fruit development remain underdeveloped. Further investment in these areas is needed to boost domestic production during off-peak periods (Abhishek Soni, 2014).

Despite the challenges, Australian-grown dragon fruit still holds a reputation for superior quality, which appeals to local consumers. This preference for Australian-grown produce is especially evident during the off-peak season (from October to April), when local fruit commands premium prices (Abhishek Soni, 2014). However, the market share of Australian-grown dragon fruit continues to shrink in the face of increasing competition from imports. Australian growers need to invest in strategies such as year-round production, targeted marketing campaigns, and improved post-harvest handling techniques to differentiate their fruit from cheaper imports.

#### 8. Current Industry Standards Worldwide

Standards for fresh produce grades are commonly employed by government agencies and private entities during the procurement process to ensure the quality of fresh fruits and vegetables (Kader, 2002). In addition to these government-mandated standards, many countries and regions also implement "voluntary standards" for various types of fresh produce. These voluntary standards allow companies to establish their own internal benchmarks, either to align with local government regulations or to meet specific market and export requirements (Barrett *et al.*, 2010). These standards are particularly useful for businesses looking to enhance the consistency and quality of their products in both domestic and international markets.

A good example of such a voluntary standard in Australia is the Pineapple Best Practice Manual developed by the Queensland Department of Primary Industries and Fisheries. This manual is used by Golden Circle Ltd., Australia as a reference for Queensland pineapple growers, guiding them in achieving the best possible quality for both fresh and processed pineapple products (Newett et al., 2018). The manual provides detailed recommendations for industry-wide quality benchmarks, such as the ideal levels of total soluble solids (TSS) in fresh Queensland-grown pineapples. For instance, the TSS level, which is a key indicator of sweetness and overall fruit quality, is recommended to be at least 16% during most times of the year, with a slightly reduced standard of 13% in winter months to account for seasonal variations. The adoption of such voluntary standards has proven to be an effective way for industries to regulate themselves and maintain high quality in their products. It also allows producers to remain flexible and

responsive to market demands while ensuring compliance with local regulations. In the global marketplace, voluntary standards are often critical for exporters, as they help to guarantee that products meet the quality expectations of international buyers and regulators, facilitating smoother trade and fostering consumer confidence.

Dragon fruit is a globally traded fruit, with production primarily concentrated in Southeast Asia and Latin America. To ensure consistency in terms of quality, safety, and market competitiveness, various countries and regions have established specific standards for the classification, grading, and handling of fresh dragon fruit. These standards dictate key parameters such as fruit size, external appearance, and post-harvest treatments. Based on available information, there are four publicly available country-specific standards (Thailand, the Philippines,

Malaysia, and Mexico), one region-specific standard (ASEAN, Southeast Asia), and one international standard (Codex Alimentarius), each addressing different aspects of fresh dragon fruit quality. Table 3 shows a detailed comparison of dragon fruit standards. The Mexican standard is not included due to the unavailability of an English version. The Codex Alimentarius Standard for Dragon Fruit (CXS 237-2003), establishes a framework for classifying and grading dragon fruit intended for fresh consumption in international trade. This standard divide dragon fruit into three classes: the Extra Class, Class I, and Class II, each reflecting different levels of quality based on external appearance and defect tolerance. Fruits categorized under the Extra Class must exhibit superior quality, with very minor superficial defects permissible, as long as they do not impact the fruit's overall aesthetic or internal quality.

**Table 10:** Comparison of different dragon fruit standards.

Standard	Classifications	Defect Tolerance	Size Range (g)	Tolerances for Size and Quality	Reference
Codex (CXS 237-2003)	Extra Class, Class I, and Class II	Extra Class: Very slight defects; Class I: Defects affecting up to 1 cm <sup>2</sup> ; Class II: Defects affecting up to 2 cm <sup>2</sup> .	110–700+ (yellow and red/white varieties)	5–10% tolerance for size and quality deviations.	Priatni & Pradita (2015)
ASEAN (ASEAN Stan 42:2015)	Extra Class, Class I, and Class II	Extra Class: Very slight defects; Class I: Defects affecting no more than 5% of the surface; Class II: Defects affecting up to 10% of the surface area.	110–700+ (red/white varieties)	5–10% tolerance for size and quality deviations.	Prosky <i>et al</i> . (1988)
Thailand (TAS 25-2015)	Extra Class, Class I, and Class II	Extra Class: No defects; Class I: Defects affecting up to 5% of the surface; Class II: Defects affecting up to 10% of the surface.	200–600+ (red-/white-fleshed varieties)	5–10% tolerance for size and quality deviations.	Putri <i>et al</i> . (2021)
Philippines (PNS/BAFPS 115:2013)	Extra Class, Class I, and Class II	Extra Class: Very slight defects; Class I: Defects affecting no more than 5% of the surface; Class II: Defects affecting up to 10% of surface area.	110–700+ (red/white varieties)	5–10% tolerance for size and quality deviations.	Ramli <i>et al</i> . (2014)
Malaysia (MS 2201: 2008)	Extra Class, Class I, and Class II	Extra Class: Very slight superficial defects; Class I: Defects affecting no more than 5% of the surface; Class II: Defects affecting up to 10% of the surface area.	110–700+ (red/white varieties)	5–10% tolerance for size and quality deviations.	Recuenco et al (2016)

Class I allows for slight defects in shape and skin, with defects covering no more than 1 cm<sup>2</sup> of the surface, while Class II permits more noticeable defects, with an allowance for defects of up to 2 cm<sup>2</sup>. The Codex standard also classifies dragon fruit based on weight, with categories ranging from 110 g to over 700 g for both red and yellow varieties. A tolerance of 5–10% is allowed for deviations in size and quality, depending on the classification of the fruit (CXS 237-2003).

Similarly, the ASEAN Standard for Dragon Fruit (ASEAN Stan 42:2015) provides guidelines for commercial dragon fruit, particularly within the Southeast Asian nations, where dragon fruit is a significant agricultural export. This standard was modified from Malaysian Standard MS 2201:2008 Fresh Pitahaya Specification (ICS: 67.080.10) and Thai Agricultural Standard TAS 25-2015. Similar to the Codex standard, the ASEAN standard categorizes dragon fruit into an Extra Class, Class I, and Class II, with specific defect

allowances corresponding to each class. Extra Class fruits must maintain near-perfect external appearance, with only slight superficial defects allowed. In Class I, minor defects in shape and skin are permissible, provided they affect no more than 5% of the fruit's surface area. Class II, which is reserved for fruit that does not meet higher quality thresholds, allows for defects on up to 10% of the surface area. The size grading in the ASEAN standard is also similar to that of the Codex standard, with weight categories ranging from 110 g to over 700 g. The

tolerance for size and quality defects ranges between 5% and 10% depending on the class (ASEAN Stan 42:2015).

The Thai Agricultural Standard (TAS 25-2015), which governs the quality of dragon fruit in Thailand, follows a similar classification system but with a few notable differences in weight categories and defect tolerances. Similar to the Codex and ASEAN standards, it defines three quality classes: an Extra Class, Class I, and Class II. The Thai standard defines slightly different size categories compared to the Codex and ASEAN standards, with dragon fruit ranging from 200 g to over 600 g in weight for red- and white-fleshed varieties. Similar to other standards, a 5–10% tolerance for size and quality is allowed depending on the class (TAS 25-2015).

The Philippine National Standard (PNS/BAFPS 115:2013) follows a similar structure to the Codex and ASEAN standards. Dragon fruits are classified into an Extra Class, Class I, and Class II based on their appearance and defect tolerances. Extra Class fruits must exhibit superior quality, while Class I allows for slight defects affecting up to 5% of the surface area. Class II permits more significant defects, covering up to 10% of the surface area. The size range is 110 g to over 700 g, and a 5–10% tolerance applies to size and quality (PNS/BAFPS 115:2013).

The Malaysian Standard (MS 2201:2008) for fresh dragon fruit is structured similarly to the other standards, categorizing fruits into an Extra Class, Class I, and Class II. Extra Class dragon fruit must be of exceptional quality and free from defects except for very slight superficial blemishes. Class I allows for minor defects, such as scratches or bruises, but limits these to a small area of the skin (up to 5%). Class II permits more pronounced defects, allowing blemishes to affect up to 10% of the surface area, similar to the Thai and Philippine standards. The size range under the Malaysian standard is also similar, with categories ranging from 110 g to over 700 g. A 5–10% tolerance for size and quality is allowed (MS 2201:2008).

# 9. Recommendation for the Establishment Dragon Fruit Standard

Establishing a robust Australian dragon fruit standard requires consideration of several factors, including bioactive phytochemicals and alignment with current global standards. The bioactive phytochemicals found in dragon fruit, such as phenolic acids, betalains, flavonoids, anthocyanins and carotenoids, offer significant health benefits. These compounds, known for their antioxidant, anti-inflammatory, and anti-microbial properties, should play a key role in differentiating Australian dragon fruit in terms of quality and nutritional value. To reflect this, the Australian standard could incorporate classifications that recognize the nutritional quality of dragon fruits in addition to external factors like

appearance and size. For example, red-fleshed varieties, which contain higher levels of antioxidants like betacyanins, could be assigned to premium categories based on their bioactive contents.

To ensure Australia remains competitive in the international market, the proposed standard should align with existing frameworks, such as the Codex Alimentarius (CXS 237-2003), ASEAN Standard (ASEAN Stan 42:2015), Thai Agricultural Standard (TAS 25-2015), and Philippine National Standard (PNS/BAFPS 115:2013), to establish classifications on size, shape, and defect tolerance. According to these standards, the Australian dragon fruit standard could use a three-tier classification system consisting of an Extra Class, Class I, and Class II. Extra Class fruits would be the highest quality, free of significant defects and imperfections, while Class I would allow for slight defects and Class II would accept more pronounced defects as long as the internal quality remains unaffected. This system would compatibility with international trade requirements, facilitating smoother export processes for Australian dragon fruit.

A unique aspect of the Australian dragon fruit standard could be the inclusion of bioactive phytochemical testing. Whereas existing standards focus predominantly on external quality, introducing optional testing for bioactive compounds like total phenolics, flavonoids, or betacyanins could elevate the status of Australian dragon fruit as a premium health product. This could create a new market segment where fruits with higher phytochemical contents are graded more favorably, promoting the health benefits of Australiangrown dragon fruit. For example, just as Brix measurements are used to indicate sweetness in some fruits, a similar system could be developed to quantify the antioxidant capacity or phytochemical concentration of dragon fruit. This would highlight the unique nutritional properties of Australian dragon fruit, especially when compared to imported varieties.

In short, an Australian dragon fruit standard should not only mirror global standards in terms of classification, size grading, and defect tolerance but also set itself apart by incorporating the nutritional and bioactive properties of the fruit. By promoting these health benefits and emphasizing sustainability, Australia could establish itself as a leader in high-quality, nutrient-rich dragon fruit production, offering a unique product in both domestic and global markets.

## 10. Strengths and Limitations of This Review

This comprehensive review consolidates current knowledge on the nutritional value, bioactive phytochemicals, therapeutic potential, and industry standards of dragon fruit, with a unique emphasis on its implications for the emerging Australian market. A key

strength lies in its holistic approach, integrating findings from diverse sources, including peer-reviewed articles, industry reports, and government publications. This enables a thorough understanding of dragon fruit's potential as a functional food and highlights gaps in global and Australian production standards.

However, the review also has limitations. The reliance on available literature may have inadvertently excluded unpublished studies or recent industry data not accessible through standard academic platforms. Additionally, while this review highlights potential bioactive benefits, the findings are primarily based on in vitro and animal studies, necessitating further validation through human clinical trials. Furthermore, the review's focus on Australian industry standards is constrained by the limited availability of local data, reflecting the nascent state of dragon fruit research and production in the region.

## **Summary and Conclusions**

In conclusion, this comprehensive review on dragon fruit highlights the significant potential of this crop in both nutritional and commercial aspects. With its origins in Mexico and northern South America, dragon fruit cultivation has expanded across tropical and subtropical regions worldwide, including Australia. The fruit is valued not only for its exotic appeal but also for its rich contents of bioactive phytochemicals such as phenols, flavonoids, and carotenoids, which offer various health benefits. Dragon fruit has been shown to possess antioxidant, anti-inflammatory, anti-cancer, and anti-microbial properties, making it an important functional food.

The global dragon fruit market is currently dominated by countries like Vietnam, China, and Indonesia, where favorable growing conditions and largescale agricultural practices have driven high productivity. However, Australia has the potential to carve out a niche market for high-quality, sustainably grown dragon fruit. The country's adherence to strict food safety regulations can further enhance its competitiveness, particularly in premium markets. For the Australian industry to grow, establishing national standards for dragon fruit production that align with international practices is crucial, especially for general quality indicators such as shape, size, and defect tolerance. By incorporating bioactive phytochemical profiles and adapting global standards to local conditions, Australia can improve the quality and marketability of its dragon fruit. This will ensure consistency in production, enhance consumer confidence, and open new opportunities for export.

Future studies are encouraged to prioritize the exploration of the influences of cultivation practices, environmental factors, and post-harvest handling techniques on the bioactive composition of dragon fruit.

Additionally, clinical trials are needed to validate the therapeutic effects of its bioactive compounds, particularly in preventing and managing chronic diseases such as diabetes, obesity, and cancer. The advancement of analytical methods for profiling and quantifying bioactive compounds will provide deeper insights into their health benefits, aiding in the establishment of bioactive-oriented industry standards. Furthermore, interdisciplinary research on innovative processing methods, such as encapsulation and preservation techniques, could enhance the stability and bioavailability of these compounds, expanding the applications of dragon fruit in functional foods and nutraceuticals. These efforts will not only strengthen the scientific understanding of dragon fruit but also support the development of industry standards that improve marketability and sustainability.

#### **Plant Protection Measures:**

#### (i) Diseases:

- Anthracnose: Pre-harvest spraying of Mancozeb 2g/lit or Carbendazim 1g/lit or Thiophanate methyl 1g/lit or Chlorothalonil 2 g/lit, 3 times at 15 days interval will control anthracnose
- **Brown Spot:** Field sanitation (collection and disposal of fallen diseased fruits, leaves and vines). Pruning vines to reduce density and thereby reducing humidity within the crop. Timely sprays with copperbased fungicides.
- **Soft rot:** Control includes pruning out dying stems and spraying with copper sulfate whitewashing before onset of the problem Timely sprays with copper-based fungicides. Copper oxy chloride (at 0.2%) can be used for managing this disease.
- Fruit rot: Maintain a weed free plan>ng and remove and discard diseased plants (i.e. stems, fruits, and flowers) promptly when symptoms occur.

## (ii) Pest:

- Aphids: Prune to avoid dense canopy, do not intercrop with alternate hosts, enhance natural enemies by incorporating natural habitats of agroforestry and flowers strips around field. Spray LambdaCyhalothrin, Cypermethrin, Amidacloprid Acetamipride, etc. are recommended, at the rate of lm/l litre of water.
- Thrips: Destroy all plant residues and volunteer plants during field preparation by turning them under while ploughing, apply mulch to reduce pupation. Spray insecticides like Deltamethrin at the rate of 1ml/1litre of water.
- Mealybug: Spray acephate 75 SP @ 1 g/l OR quinalphos 25 EC 2 ml/l or chlorpyriphos 25 EC 2ml/l or profenophos 50 EC @ 2 ml or thiodicarb 75 WP 2gm/l

#### Availability of data and material

All relevant data and material are presented in the Research

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#### **Ethics declarations**

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## **References**

- Adetuyi, F. O., & Ibrahim, T. A. (2014). Effect of fermentation time on the phenolic, flavonoid and vitamin C contents and antioxidant activities of okra (*Abelmoschus esculentus*) seeds. *Nigerian Food Journal*, 32, 128–137. https://doi.org/10.1016/S0189-7241(15)30128-4
- Adinortey, M.B., Agbeko, R., Boison, D., Ekloh, W., Kuatsienu, L.E., Biney, E.E., Affum, O.O., Kwarteng, J., Nyarko, A.K. Phytomedicines used for diabetes mellitus in Ghana: A systematic search and review of preclinical and clinical evidence. *Evid. Based Complement. Altern. Med.* 2019, 2019, 6021209.
- Ahmadi, A., Shadboorestan, A., Nabavi, S., Setzer, W., Nabavi, S. The role of hesperidin in cell signal transduction pathway for the prevention or treatment of cancer. *Curr. Med. Chem.* 2015, 22, 3462–3471.
- Ali Jaafar, R., Abdul Rahm, A. R. B., Che Mahmod, N. Z., & Vasudevan, R. (2009). Proximate analysis of dragon fruit (Hylecereus polyhizus). *American Journal of Applied Sciences*, 6, 1341–1346.
- Al-Mekhlafi, N.A., Mediani, A., Ismail, N.H., Abas, F., Dymerski, T., Lubinska-Szczygeł, M., Vearasilp, S.,

- Gorinstein, S. Metabolomic and antioxidant properties of different varieties and origins of Dragon fruit. *Microchem. J.* 2021, *160*, 105687.
- Ariffin, A., Bakar, J., Tan, C., Rahman, R., Karim, R., & Loi, C. (2009). Essential fatty acids of pitaya (dragon fruit) seed oil. *Food Chemistry*, 114, 561–564. https://doi.org/10.1016/j.foodchem.2008.09.108
- Ariffin, A.A., Bakar, J., Tan, C.P., Rahman, R.A., Karim, R., Loi, C.C. Essential fatty acids of pitaya (dragon fruit) seed oil. *Food Chem.* 2009, 114, 561–564.
- Arivalagan, M., Karunakaran, G., Roy, T., Dinsha, M., Sindhu, B., Shilpashree, V., Satisha, G., Shivashankara, K. Biochemical and nutritional characterization of dragon fruit (*Hylocereus* species). Food Chem. 2021, 353, 129426
- Attar, Ş.H., Gündeşli, M.A., Urün, I., Kafkas, S., Kafkas, N.E., Ercisli, S., Ge, C., Mlcek, J., Adamkova, A. Nutritional analysis of red-purple and white-fleshed pitaya (*Hylocereus*) species. *Molecules* 2022, 27, 808.
- Attoe, E. L., & Elbe, J. H. V. O. N. (1981). Photochemial degradation of betanine & I selected anthocyanins pigment analysis. *Journal of Food Science*, 46, 1934– 1937
- Attoe, E. L., & Von Elbe, J. H. (1982). Degradation kinetics of betanin in solutions as influenced by oxygen. *Journal of Agricultural and Food Chemistry*, 30, 708–712.
- Aued-pimentel, S., Takemoto, E., Antoniassi, R., & Gastaldo Badolato, E. S. (2006). Composition of tocopherols in sesame seed oil: An indicative of adulteration. *Grasas Y Aceites*, 57, 205–210.
- Azeredo, H. M. C. (2009). Betalains: Properties, sources, applications, and stability A review. *International Journal of Food Science & Technology*, 44, 2365–2376. https://doi.org/10.1111/j.1365-2621.2007.01668.x
- Baliga, M.S., Shivashankara, A.R., Haniadka, R., Dsouza, J., Bhat, H.P. Phytochemistry, nutritional and pharmacological properties of *Artocarpus heterophyllus* Lam (jackfruit): A review. *Food Res. Int.* 2011, 44, 1800– 1811.
- Barrett, D.M., Beaulieu, J.C., Shewfelt, R. Color, flavor, texture, and nutritional quality of fresh-cut fruits and vegetables: Desirable levels, instrumental and sensory measurement, and the effects of processing. *Crit. Rev. Food Sci. Nutr.* 2010, 50, 369–389.
- Baydar, N. G., Özkan, G., & Çetin, E. S. (2007). Characterization of grape seed and pomace oil extracts. *Grasas Y Aceites*, 58, 29–33.
- Bellec, F. L., & Vaillant, F. (2011). Pitahaya (pitaya) (Hylocereus spp.). In E. M. Yahia, (Ed.), *Postharvest biology and technology of tropical and subtropical fruits* (pp. 247–273). Woodhead Publishing Limited. https://doi.org/10.1533/9780857092618.247
- Charoensiri, R., Kongkachuichai, R., Suknicom, S., Sungpuag, P. Beta-carotene, lycopene, and alpha-tocopherol contents of selected Thai fruits. *Food Chem.* 2009, *113*, 202–207.
- Chen, S.-Y., Xu, C.-Y., Mazhar, M.S., Naiker, M. Nutritional Value and Therapeutic Benefits of Dragon Fruit: A Comprehensive Review with Implications for Establishing Australian Industry Standards. *Molecules* 2024, 29, 5676. https://doi.org/10.3390/molecules29235676
- Chen, Z., Zhong, B., Barrow, C.J., Dunshea, F.R., Suleria, H.A.
  Identification of phenolic compounds in Australian grown
  dragon fruits by LC-ESI-QTOF-MS/MS and

- determination of their antioxidant potential. Arab. J. Chem. 2021, 14, 103151.
- Chlebicz-Wójcik, A., Śliżewska, K. Probiotics, prebiotics, and synbiotics in the irritable bowel syndrome treatment: A review. *Biomolecules* 2021, 11, 1154.
- Choo, K. Y., Kho, C., Ong, Y. Y., Ho, C. W., & Lim, L. H. (2018). Fermentation of red dragon fruit (Hylocereus polyrhizus) for betalains concentration. International Food Research Journal, 25, 2539–2546.
- Craig, L. S., Russell, L. C., Cindy, L. S., & Timothy, P. C. (2000). Dietary Stearic Acid Reduces Cholesterol Absorption and Increases Endogenous Cholesterol Excretion in Hamsters Fed Cereal-Based Diets. *Journal of Nutrition*, 130, 1232–1238.
- Crizel, T. M., Araujo, R. R., Rios, A. O., Rech, R., & Flôres, S. H. (2014). Orange fiber as a novel fat replacer in lemon ice cream. Food Science and Technology (Campinas), 34, 332–340.
- Dasaesamoh, R., Youravong, W., Wichienchot, S. Digestibility, fecal fermentation and anti-cancer of dragon fruit oligosaccharides. *Int. Food Res. J.* 2016, *23*, 2581–2587.
- Dayal, V., Kumar, S., Boopathi, T., Dutta, S., Saha, S., Singh, S. Dragon fruit: A potential source for nutritional security. *Indian Hortic.* 2018, *63*, 32–35.
- De Brabandere, A. G., & De Baerdemaeker, J. G. (1999). Effects of process conditions on the pH development during yogurt fermentation. *Journal of Food Engineering*, 41, 221–227.
- Demo, P., Kuria, P., Nyende, A., & Kahangi, E. (2008). Table sugar as an alternative low-cost medium component for in vitro micro-propagation of potato (*Solanum tuberosum* L.). *African Journal of Biotechnology*, 7, 2578–2584.
- Diez, J., & Iglesias, P. (2003). The role of the novel adipocytederived hormone adiponectin in human disease. *European Journal of Endocrinology*, 148, 293–300.
- Dimero, F. N., & Tepora, T. F. (2018). Processing and development of dragon fruit wine. *International Journal* of Agriculture Environment and Biotechnology, 3, 1943– 1947.
- Ding, X., Zhu, X., Zheng, W., Li, F., Xiao, S., Duan, X. BTH treatment delays the senescence of postharvest pitaya fruit in relation to enhancing antioxidant system and phenylpropanoid pathway. *Foods* 2021, *10*, 846.
- Divakaran, D., Lakkakula, J.R., Thakur, M., Kumawat, M.K., Srivastava, R. Dragon fruit extract capped gold nanoparticles: Synthesis and their differential cytotoxicity effect on breast cancer cells. *Mater. Lett.* 2019, 236, 498– 502.
- Ehsan, S.D. Determination of pitaya seeds as a natural antioxidant and source of essential fatty acids. *Int. Food Res. J.* 2010, *17*, 1003–1010.
- Esatbeyoglu, T., Wagner, A. E., Schini-Kerth, V. B., & Rimbach, G. (2015). Betanin-A food colorant with biological activity. *Molecular Nutrition & Food Research*, 59, 36–47.
- Fadahunsi, I. F., & Sanni, A. I. (2010). Chemical and Biochemical Changes in Bambara Nut (Voandzeia subterranean) during Fermentation to 'Tempeh'. Electronic Journal of Environmental. *Agriculture and Food Chemistry*, 9(2), 275–283.
- Felix, E., Clara, O., & Vincent, A. O. (2014). Thermodynamic characterization of *Saccharomyces cerevisiae* catalyzed

- fermentation of cane sugar. Open Journal of Physical Chemistry, 4, 21–25.
- Ferrari, C. C., Germer, S. P. M., & de Aguirre, J. M. (2012). Effects of spray-drying conditions on the physicochemical properties of blackberry powder. *Drying Technology*, 30, 154–163, https://doi.org/10.1080/07373937.2011.628429
- Fidrianny, I., Ilham, N., Hartati, R. Antioxidant profile and phytochemical content of different parts of super red dragon fruit (*Hylocereus costaricensis*) collected from West Java-Indonesia. *Asian J. Pharm. Clin. Res.* 2017, 10, 290–294.
- Figuerola, F., Hurtado, M. L., Estevez, A. M., Choffelle, I., & Asenjo, F. (2005). Fiber concentrates from apple pomace and citrus peels as potential fiber sources for food enrichment. *Food Chemistry*, 91, 395–401.
- Frick, E. M., & Strader, L. C. (2018). Roles for IBA-derived auxin in plant development. *Journal of Experimental Botany*, 69, 169–177. https://doi.org/10.1093/jxb/erx298
- Gengatharan, A., Dykes, G. A., & Choo, W. S. (2016). Stability of betacyanin from red pitahaya (*Hylocereus polyrhizus*) and its potential application as a natural colourant in milk. *International Journal of Food Science & Technology*, 51, 427–434. https://doi.org/10.1111/jifs.12999
- Gong, X., Ma, L., Li, L., Yuan, Y., Peng, S., Lin, M. Analysis of aroma compounds of pitaya fruit wine. *IOP Conf. Ser. Earth Environ. Sci.* 2017, 100, 012125.
- Gülcin, I. Antioxidant activity of food constituents: An overview. Arch. Toxicol. 2012, 86, 345–391.
- Harahap, N. S., & Amelia, R. (2019). Red dragon fruit (Hylocereus polyrhizus) extract decreases lactic acid level and creatine kinase activity in rats receiving heavy physical exercise. Open access Macedonian Journal of Medical Sciences, 7, 2232–2235.
- Harahap, N.S., Amelia, R. Red dragon fruit (*Hylocereus polyrhizus*) extract decreases lactic acid level and creatine kinase activity in rats receiving heavy physical exercise. *Open Access Maced. J. Med. Sci.* 2019, 7, 2232.
- Hendra, R., Khodijah, R., Putri, R., Amalia, R., Haryani, Y., Teruna, H.Y., Abdulah, R. Cytotoxicity and antiplasmodial properties of different *Hylocereus polyrhizus* peel extracts. *Med. Sci. Monit. Basic Res.* 2021, 27, e931118-1.
- Herbach, K. M., & Stintzing, F. C. (2004). Thermal degradation of betacyanins in juices from purple pitaya *Hylocereus polyrhizus* (Weber) Britton & Rose monitored by high-performance liquid chromatography—Tandem mass spectometric analyses. *European Food Research and Technology*, 377–385. https://doi.org/10.1007/s00217-004-0948-8
- Herbach, K. M., Maier, C., Stintzing, F. C., & Carle, R. (2007). Effects of processing and storage on juice colour and betacyanin stability of purple pitaya (*Hylocereus polyrhizus*) juice. *European Food Research and Technology*, 224, 649–658. https://doi.org/10.1007/s00217-006-0354-5
- Herbach, K. M., Rohe, M., Stintzing, F. C., & Carle, R. (2006). Structural and chromatic stability of purple pitaya (*Hylocereus polyrhizus* Weber Britton & Rose) betacyanins as affected by the juice matrix and selected additives. *Food Research International*, 39, 667–677. https://doi.org/10.1016/j.foodres.2006.01.004
- Ho, L.-H., Abdul Latif, N.W.b. Nutritional composition, physical properties, and sensory evaluation of cookies

- prepared from wheat flour and pitaya (*Hylocereus* undatus) peel flour blends. *Cogent Food Agric*. 2016, 2, 1136369.
- Hossain, M. N. (2012). Quality comparison and acceptability of yoghurt with different fruit juices. *Journal of Food Processing & Technology*, 3, 8–12. https://doi.org/ 10.4172/2157-7110.1000171
- Huang, Y., Brennan, M.A., Kasapis, S., Richardson, S.J., Brennan, C.S. Maturation process, nutritional profile, bioactivities and utilisation in food products of red pitaya fruits: A Review. *Foods* 2021, 10, 2862.
- Ibrahim, S.R.M., Mohamed, G.A., Khedr, A.I.M., Zayed, M.F., El-Kholy, A.A.E.S. Genus *Hylocereus*: Beneficial phytochemicals, nutritional importance, and biological relevance A review. *J. Food Biochem.* 2018, *42*, e12491.
- Islam, M. Z., Khan, M. T. H., Hoque, M. M., & Rahman, M. M. (2012). Studies on the processing and preservation of dragon fruit (*Hylocereus undatus*) jelly. *The Agriculturists*, 10, 29–35.
- Islam, M., Khan, M., Hoque, M., Rahman, M. Studies on the processing and preservation of dragon fruit (*Hylocereus undatus*) jelly. *Agriculturists* 2012, *10*, 29–35.
- Ismail, N., Ramli, N., Hani, N., & Meon, Z. (2012). Extraction and characterization of pectin from dragon fruit (*Hylocereus polyrhizus*) using various extraction conditions. *Sains Malaysiana*, 41, 41–45.
- Jalgaonkar, K., Mahawar, M. K., Bibwe, B., & Kannaujia, P. (2022). Postharvest profile, processing and waste utilization of dragon fruit (Hylocereus Spp.): A review. Food Reviews International, 38, 733–759.
- Jayakumar, R., & Kanthimathi, M. S. (2011). Inhibitory effects of fruit extracts on nitric oxide induced proliferation in MCF-7 cells. *Food Chemistry*, 126, 956–960.
- Jayakumar, R., Kanthimathi, M. Inhibitory effects of fruit extracts on nitric oxide-induced proliferation in MCF-7 cells. Food Chem. 2011, 126, 956–960.
- Jeong, J. H., Jung, H., Lee, S. R., Lee, H. J., Hwang, K. T., & Kim, T. Y. (2010). Antioxidant, anti-proliferative and anti-inflammatory activities of the extracts from black raspberry fruits and wine. *Food Chemistry*, 123, 338–344.
- Jiang, X., Lu, Y., & Liu, S. Q. (2020). Effects of different yeasts on physicochemical and oenological properties of red dragon fruit wine fermented with *Saccharomyces cerevisiae*, Torulaspora delbrueckii and Lachancea thermotolerans. *Microorganisms*, 8, 315.
- Kamairudin, N., Gani, S.S.A., Masoumi, H.R.F., Hashim, P. Optimization of natural lipstick formulation based on pitaya (*Hylocereus polyrhizus*) seed oil using D-optimal mixture experimental design. *Molecules* 2014, 19, 16672–16683.
- Kashyap, D. R., Vohra, P. K., Chopra, S., & Tewari, R. (2001).
  Applications of pectinases in the commercial sector: A review. *Bioresource Technology*, 77, 215–227.
- Kaul, K., Tarr, J.M., Ahmad, S.I., Kohner, E.M., Chibber, R. Introduction to diabetes mellitus. Adv. Exp. Med. Biol. 2012, 771, 1–11.
- Khalili, R.M.A., Norhayati, A., Rokiah, M., Asmah, R., Nasir, M.M., Muskinah, M.S. Proximate composition and selected mineral determination in organically grown red pitaya (*Hylocereus* sp.). J. Trop. Agric. Food Sci. 2006, 34, 269.
- Kim, H., Choi, H.K., Moon, J.Y., Kim, Y.S., Mosaddik, A., Cho, S.K. Comparative antioxidant and antiproliferative

- activities of red and white pitayas and their correlation with flavonoid and polyphenol content. *J. Food Sci.* 2011, 76, C38–C45.
- Kirsten, M. H., Florian, C. S., & Reinhold, C. (2006). Betalain stability and degradation-structural and chromatic aspects. *Journal of Food Science*, 71, 41–50.
- Krichene, D., Allalout, A., Mancebo-campos, V., Salvador, M. D., Zarrouk, M., & Fregapane, G. (2010). Stability of virgin olive oil and behaviour of its natural antioxidants under medium temperature accelerated storage conditions. Food Chemistry, 121, 171–177. https://doi.org/10.1016/j.foodchem.2009.12.026
- Kumar, S.B., Issac, R., Prabha, M.L. Functional and health-promoting bioactivities of dragon fruit. *Drug Invent. Today* 2018, 10, 3307–3310.
- Kumorkiewicz-Jamro, A., Świergosz, T., Sutor, K., Spórna-Kucab, A., Wybraniec, S. Multi-colored shades of betalains: Recent advances in betacyanin chemistry. *Nat. Prod. Rep.* 2021, *38*, 2315–2346.
- Le Bellec, F., Vaillant, F. Pitahaya (Pitaya) (*Hylocereus* spp.). In *Postharvest Biology and Technology of Tropical and Subtropical Fruits*, Elsevier: Amsterdam, The Netherlands, 2011, pp. 247–273.
- Le, N.L. Functional compounds in dragon fruit peels and their potential health benefits: A review. *Int. J. Food Sci. Technol.* 2022, *57*, 2571–2580.
- Le, T.-L., Huynh, N., & Quintela-Alonso, P. (2021). Dragon fruit: A review of health benefits and nutrients and its sustainable development under climate changes in Vietnam. Czech Journal of Food Sciences, 39(2), 71–94.
- Le, T.-L., Huynh, N., Quintela-Alonso, P. Dragon fruit: A review of health benefits and nutrients and its sustainable development under climate changes in Vietnam. *Czech J. Food Sci.* 2021, 39, 71–94.
- Lee, K., Wu, T., & Siow, L. (2013). Spray drying of red (*Hylocereus polyrhizus*) and white (*Hylocereus undatus*) dragon fruit juices: Physicochemical and antioxidant properties of the powder. *International Journal of Food Science and Technology*, 48, 2391–2399. https://doi.org/10.1111/jifs.12230
- Lemcke-noroja, M., Kamal-eldin, A., Appelqvist, L. Å, Dimberg, L. H., & Vessby, B. (2001). Corn and sesame oils increase serum gamma-tocopherol concentrations in healthy Swedish women. *The American Society for Nutritional Sciences*, 131, 1195–1201.
- Leuenberger, H., & Dass Rohera, B. (1986). Fundamentals of powder compression. I. The compactibility and compressibility of pharmaceutical powders. *Pharmaceutical Research*, 3, 12–22.
- Li, F., Li, S., Li, H. B., Deng, G. F., Ling, W. H., Wu, S., Xu, X. R., & Chen, F. (2013). Antiproliferative activity of peels, pulps and seeds of 61 fruits. *Journal of Functional Foods*, 5, 1298–1309. https://doi.org/10.1016/j.jff.2013.04.016
- Liaotrakoon, W., & Clercq, N. D. (2013). Impact of thermal treatment on physicochemical, antioxidative and rheological properties of white-flesh and red-flesh dragon fruit (Hylocereus spp.) purees. *Food and Bioprocess Technology*, 6, 416–430. https://doi.org/10.1007/s11947-011-0722-4
- Liaotrakoon, W., Clercq, N. D., Hoed, V. V., & Dewettinck, K. (2013). Dragon fruit (Hylocereus spp.) seed oils: Their characterization and stability under storage conditions.

- Journal of the American Oil Chemists' Society, 90, 207–215. https://doi.org/10.1007/s11746-012-2151-6
- Liaotrakoon, W., Van Buggenhout, S., Christiaens, S., Houben,
  K., De Clercq, N., Dewettinck, K., & Hendrickx, M. E.
  (2013). An explorative study on the cell wall polysaccharides in the pulp and peel of dragon fruits
  (Hylocereus spp.). European Food Research and Technology, 237, 341–351. https://doi.org/10.1007/s00217-013-1997-7
- Lim, H.K., Tan, C.P., Karim, R., Ariffin, A.A., Bakar, J. Chemical composition and DSC thermal properties of two species of *Hylocereus cacti* seed oil: *Hylocereus undatus* and *Hylocereus polyrhizus*. Food Chem. 2010, 119, 1326– 1331.
- Lim, Y.Y., Lim, T.T., Tee, J.J. Antioxidant properties of several tropical fruits: A comparative study. *Food Chem.* 2007, 103, 1003–1008.
- Liu, B., Ye, C., Geng, L., & Zhou, W. (2019). Betacyanins in dragon fruit peels: the kinetic models of their degradation under different treatment conditions. *International Journal of Food Science & Technology*, 54, 2949–2962. https://doi.org/10.1111/ijfs.14206
- Lourens-hattingh, A., & Viljoen, B. C. (2001). Yogurt as probiotic carrier food. *International Dairy Journal*, 11, 1– 17.
- Lu, Y., Huang, D., Lee, P. R., & Liu, S. Q. (2015). Effects of cofermentation and sequential inoculation of Saccharomyces bayanus and Torulaspora delbruckii on durian wine composition. *International Journal of Food Science & Technology*, 50, 2653–2663. https://doi.org/ 10.1111/ijfs.12937
- Luo, H., Cai, Y., Peng, Z., Liu, T., Yang, S. Chemical composition and in vitro evaluation of the cytotoxic and antioxidant activities of supercritical carbon dioxide extracts of pitaya (dragon fruit) peel. *Chem. Cent. J.* 2014, 8, 1–7.
- Lwin, T., Myint, C.Y., Win, H.H., Oo, W.W., Chit, K. Formulation and evaluation of lipstick with betacyanin pigment of *Hylocereus polyrhizus* (Red Dragon Fruit). *J. Cosmet. Dermatol. Sci. Appl.* 2020, *10*, 212–224.
- Lyu, J., Jian, yong Y., Feng Bi, J., Yan Zhou, L., Zhou, M. (2017). Hot air drying and freeze-drying pre-treatments coupled to explosion pu ffi ng drying in terms of quality attributes of mango, pitaya, and papaya fruit chips. *Journal of Food Processing and Preservation*, 41(6), 1–10. https://doi.org/10.1111/jfpp.13300
- Mahattanatawee, K., Manthey, J. A., Luzio, G., Talcott, S. T., Goodner, K., & Baldwin, E. A. (2006). Total antioxidant activity and fiber content of select Florida-grown tropical fruits. *Journal of Agricultural and Food Chemistry*, 54, 7355–7363.
- Mahayothee, B., Komonsing, N., Khuwijitjaru, P., Nagle, M., Müller, J. Influence of drying conditions on colour, betacyanin content and antioxidant capacities in dried redfleshed dragon fruit (*Hylocereus polyrhizus*). *Int. J. Food Sci. Technol.* 2019, *54*, 460–470.
- Mahdi, M. A., Mohammed, M. T., Jassim, A. M. N., & Mohammed, A. I. (2018). Phytochemical content and antioxidant activity of Hylocereus undatus and study of toxicity and the ability of wound treatment. *Plant Archives*, 18(2), 2672–2680.
- Manh, B., Nguyen, N., & Pirak, T. (2019). Physicochemical properties and antioxidant activities of white dragon fruit

- peel pectin extracted with conventional and ultrasound-assisted extraction. *Cogent Food Agriculture*, 5, 1–13. https://doi.org/10.1080/23311932.2019.1633076
- Manihuruk, F.M., Suryati, T., Arief, I.I. Effectiveness of the red dragon fruit (*Hylocereus polyrhizus*) peel extract as the colorant, antioxidant, and antimicrobial on beef sausage. *Media Peternak*. 2017, 40, 47–54.
- Mazlina, M. K. S., & Taip, F. S. (2009). Effects of commercial pectinases application on selected properties of red pitaya juice. *Journal of Food Process Engineering*, 34, 1523– 1534. https://doi.org/10.1111/j.1745-4530.2009.00388.x
- Mazlina, M. K. S., Taip, F. S., & Abdullah, A. G. L. (2008). Response surface optimization for clarification of white pitaya juice using a commercial enzyme. *Journal of Food Process Engineering*, 33, 333–347. https://doi.org/10.1111/j.1745-4530.2008.00277.x
- Mercado-silva, E. M. (2018). Pitaya—Hylocereus undatus (Haw). In S. Rodrigues, E. Oliveira Silva, & E. S. Brito (Eds.), Exotic fruits reference guide. Elsevier Inc. https://doi.org/10.1016/B978-0-12-803138-4.00045-9
- Mercado-Silva, E.M. Pitaya—*Hylocereus undatus* (haw). In *Exotic Fruits*, Academic Press: Cambridge, MA, USA, 2018, pp. 339–349.
- Mizrahi, Y. (2014). Vine-cacti pitayas—The new crops of the world. *Revista Brasileira de Fruticultura*, 1, 124–138.
- Mizrahi, Y., Nerd, A., & Nobel, P. S. (1997). Cacti as crops. In J. Janick (Ed.), *Horticultural reviews* 18, 291–319. John Wiley & Sons.
- Mohammed, A. T. I., Chowdhury, M. A. U., Uddin, M. E., Rahman, M. M., Habib, M. R., Uddin, M. G. M., & Rahman, M. A. (2013). Protective Effect of Methanolic Extract of *Hylocereus polyrhizus* Fruits on Carbon Tetra Chloride-Induced Hepatotoxicity in Rat. *European Journal of Medicinal Plants*, 3(4), 500–507.
- Molina, R., Clemente, E., Scapim, M. R. S., & Vagula, J. M. (2014). Physical evaluation and hygroscopic behavior of dragon fruit (*Hylocereus undatus*) lyophilized pulp powder. *Drying Technology*, 32, 2005–2011. https://doi.org/10.1080/07373937.2014.929587
- Montoya-Arroyo, A., Schweiggert, R.M., Pineda-Castro, M.-L., Sramek, M., Kohlus, R., Carle, R., Esquivel, P. Characterization of cell wall polysaccharides of purple pitaya (*Hylocereus* sp.) pericarp. *Food Hydrocoll*. 2014, 35, 557–564.
- Moo-Huchin, V.M., Gonzlez-Aguilar, G.A., Moo-Huchin, M., Ortiz-V, E. Carotenoid composition and antioxidant activity of extracts from tropical fruits. *Chiang Mai J. Sci.* 2017, *44*, 605–616.
- Morais, S.G.G., Borges, G.d.S.C., dos Santos Lima, M., Martín-Belloso, O., Magnani, M. Effects of probiotics on the content and bioaccessibility of phenolic compounds in red pitaya pulp. Food Res. Int. 2019, 126, 108681.
- Naseer, S., Hussain, S., Abid, A. Betalain as a Food Colorant: Its Sources, Chemistry and Health Benefits: Chemistry of Betalain and its Role as Food Colorant. *Proc. Pak. Acad. Sci. B Life Environ. Sci.* 2019, *56*, 1–8.
- Nizamlıoğlu, N.M., Ünver, A., Kadakal, Ç. Mineral Content of Pitaya (*Hylocereus polyrhizus* and *Hylocereus* undatus) Seeds Grown in Turkey. *Erwerbs-Obstbau* 2021, *63*, 209–213.
- Nurmahani, M., Osman, A., Hamid, A.A., Ghazali, F.M., Dek, M. Antibacterial property of *Hylocereus polyrhizus* and

- Hylocereus undatus peel extracts. Int. Food Res. J. 2012, 19, 77–84.
- Nurul, S., & Asmah, R. (2014). Variability in nutritional composition and phytochemical properties of red pitaya (Hylocereus polyrhizus) from Malaysia and Australia. The International Food Research Journal, 21, 1689–1697.
- Omidizadeh, A., Yusof, R. M., Roohinejad, S., Ismail, A., Bakar, M. Z. A., & Bekhit, A. E.-D. A. (2014). Anti-diabetic activity of red pitaya (*Hylocereus polyrhizus*) fruit. *RSC Advances*, 4(108), 62978–86.
- Omidizadeh, A., Yusof, R.M., Roohinejad, S., Ismail, A., Bakar, M.Z.A., Bekhit, A.E.-D.A. Anti-diabetic activity of red pitaya (*Hylocereus polyrhizus*) fruit. *RSC Adv.* 2014, 4, 62978–62986.
- Ozmihci, S., & Kargi, F. (2007). Effects of feed sugar concentration on continuous ethanol fermentation of cheese whey powder solution (CWP). *Enzyme and Microbial Technology*, 41, 876–880.
- Padmavathy, K., Sivakumari, K., Karthika, S., Rajesh, S., Ashok, K. Phytochemical profiling and anticancer activity of dragon fruit *Hylocereus undatus* extracts against human hepatocellular carcinoma cancer (hepg-2) cells. *Int. J. Pharm. Sci. Res.* 2021, 12, 2770–2778.
- Paśko, P., Galanty, A., Zagrodzki, P., Luksirikul, P., Barasch, D., Nemirovski, A., Gorinstein, S. Dragon fruits as a reservoir of natural polyphenolics with chemopreventive properties. *Molecules* 2021, 26, 2158.
- Penna, A. L. B., Sivieri, K., & Oliveira, M. N. (2001). Relation between quality and rheological properties of lactic beverages. *Journal of Food Engineering*, 49, 7–13.
- Phongtongpasuk, S., Poadang, S., Yongvanich, N. Environmental-friendly method for synthesis of silver nanoparticles from dragon fruit peel extract and their antibacterial activities. *Energy Procedia* 2016, 89, 239–247
- Poolsup, N., Suksomboon, N., & Paw, N. J. (2017). Effect of dragon fruit on glycemic control in prediabetes and type 2 diabetes: A systematic review and meta-analysis. *PLoS One*, 12(9), 1–12.
- Poolsup, N., Suksomboon, N., Paw, N.J. Effect of dragon fruit on glycemic control in prediabetes and type 2 diabetes: A systematic review and meta-analysis. *PLoS ONE* 2017, 12, e0184577.
- Priatni, S., & Pradita, A. (2015). Stability study of betacyanin extract from red dragon fruit (*Hylocereus polyrhizus*) peels. *Procedia Chemistry*, 16, 438–444. https://doi.org/10.1016/j.proche.2015.12.076
- Prosky, L., Asp, N.-G., Schweizer, T., DeVries, J., & Furda, I. (1988). Determination of insoluble, soluble, and total dietary fiber in foods and food products: interlaboratory study. *Journal Association of Official Analytical Chemists*, 71(5), 1017–1023.
- Putri, M.D., Wiboworini, B., Dirgahayu, P. Red dragon fruit juice in reducing ros levels and insulin resistance in rats with type 2 diabetes mellitus model. *J. Gizi Indones.* (*Indones. J. Nutr.*) 2021, *10*, 6–14.
- Ramli, N. S., Ismail, P., & Rahmat, A. (2014). Influence of Conventional and Ultrasonic-Assisted Extraction on Phenolic Contents, Betacyanin Contents, and Antioxidant Capacity of Red Dragon Fruit (Hylocereus polyrhizus). The Scientific World Journal, 2014, 1–7.
- Recuenco, M.C., Lacsamana, M.S., Hurtada, W.A., Sabularse, V.C. Total phenolic and total flavonoid contents of

- selected fruits in the Philippines. Philipp. J. Sci. 2016, 145, 275–281.
- Ritarwan, K., Nerdy, N. Antibacterial activity of red dragon fruit leaves extract and white dragon fruit leaves extract against meningitis bacterial. *Orient. J. Chem.* 2018, 34, 2534.
- Rodriguez, E.B., Vidallon, M.L.P., Mendoza, D.J.R., Reyes, C.T. Health-promoting bioactivities of betalains from red dragon fruit (*Hylocereus polyrhizus* (Weber) Britton and Rose) peels as affected by carbohydrate encapsulation. *J. Sci. Food Agric.* 2016, *96*, 4679–4689.
- Rokiah, M. Y. H., Asmah, R., Muskinah, M. S., Manaf, A. A., Khalili, R.M.A., & Norhayati, A. (2009). Hypocholesterolemic effect of red pitaya (*Hylocereus* sp.) on hypercholesterolemia induced rats. *The International Food Research Journal*, 16, 431–440.
- Rui, H., Zhang, L., Li, Z., & Pan, Y. (2009). Extraction and characteristics of seed kernel oil from white pitaya. *Journal of Food Engineering*, 93, 482–486. https://doi. org/10.1016/j.jfoodeng.2009.02.016
- Saenjum, C., Pattananandecha, T., Nakagawa, K. Antioxidative and anti-inflammatory phytochemicals and related stable paramagnetic species in different parts of dragon fruit. *Molecules* 2021, 26, 3565.
- Safira, A., Savitri, S.L., Putri, A.R.B., Hamonangan, J.M., Safinda, B., Solikhah, T.I., Khairullah, A.R., Puspitarani, G.A. Review on the pharmacological and health aspects of *Hylocereus* or Pitaya: An update. *J. Drug Deliv. Ther*. 2021, 11, 297–303.
- Setiawan, N. A., Shintawati, R., & Priyandoko, D. (2018). The role of red dragon fruit peel (*Hylocereus polyrhizus*) to improvement blood lipid levels of hyperlipidaemia male mice. *Journal of Physics: Conference Series*, 1013(1), 012167
- Shaaruddin, S., Ghazali, H. M., Hamed Mirhosseini, S., & Muhammad, K. (2017). Stability of betanin in pitaya powder and confection as affected by resistant maltodextrin. *LWT*, 84, 129–134. https://doi.org/10.1016/j.lwt.2017.05.031
- Song, H., Chu, Q., Yan, F., Yang, Y., Han, W., & Zheng, X. (2016). Red pitaya betacyanins protects from diet-induced obesity, liver steatosis and insulin resistance in association with modulation of gut microbiota in mice. *Journal of Gastroenterology and Hepatology*, 31, 1462–1469. https://doi.org/10.1111/jgh.13278
- Song, H., Chu, Q., Yan, F., Yang, Y., Han, W., Zheng, X. Red pitaya betacyanins protects from diet-induced obesity, liver steatosis and insulin resistance in association with modulation of gut microbiota in mice. *J. Gastroenterol. Hepatol.* 2016, *31*, 1462–1469.
- Srinivasulu, C., Ramgopal, M., Ramanjaneyulu, G., Anuradha, C., Kumar, C.S. Syringic acid (SA) A review of its occurrence, biosynthesis, pharmacological and industrial importance. *Biomed. Pharmacother.* 2018, 108, 547–557.
- Stintzing, F. C., & Carle, R. (2007). Betalains Emerging prospects for food scientists. *Trends in Food Science & Technology*, 18, 514–525. https://doi.org/10.1016/j.tifs. 2007.04.012
- Strack, D., Vogt, T., & Schliemann, W. (2003). Recent advances in betalain research. *Phytochemistry*, 62, 247– 269.
- Sudha, K., Baskaran, D., Ramasamy, D., Siddharth, M. Evaluation of functional properties of *Hylocereus undatus*

- (White dragon fruit). *Int. J. Agric. Sci. Res.* 2017, 7, 451–456.
- Swami, S.B., Thakor, N., Haldankar, P., Kalse, S. Jackfruit and its many functional components as related to human health: A review. *Compr. Rev. Food Sci. Food Saf.* 2012, 11, 565–576.
- Tallei, T., Tumilaar, S., Lombogia, L., Adam, A., Sakib, S., Emran, T., Idroes, R. Potential of betacyanin as inhibitor of SARS-CoV-2 revealed by molecular docking study. *IOP Conf. Ser. Earth Environ. Sci.* 2021, 711, 012028.
- Tenore, G.C., Novellino, E., Basile, A. Nutraceutical potential and antioxidant benefits of red pitaya (*Hylocereus polyrhizus*) extracts. *J. Funct. Foods* 2012, *4*, 129–136.
- Toma, M., Vinatoru, M., Paniwnyk, L., & Mason, T. J. (2001). Investigation of the effects of ultrasound on vegetal tissues during solvent extraction. *Ultrasonics Sonochemistry*, 8, 137–142.
- Tongkham, N., Juntasalay, B., Lasunon, P., & Sengkhamparn, N. (2017). Dragon fruit peel pectin: Microwave-assisted extraction and fuzzy assessment. Agriculture and Natural Resources, 51, 262–267.
- Torres, R. C., Yumang, R. M. G., & Jose, C. K. F. (2020). Physicochemical properties and stability of microencapsulated betacyanin pigments from red dragon fruit peels and flesh. *Open Journal of Pharmaceutical Science and Research*, 2, 141–148.
- Tsai, Y., Lin, C.-G., Chen, W.-L., Huang, Y.-C., Chen, C.-Y., Huang, K.-F., Yang, C.-H. Evaluation of the antioxidant and wound-healing properties of extracts from different parts of *Hylocereus polyrhizus*. *Agronomy* 2019, *9*, 27.
- Tze, N. L., Han, C. P., Yusof, Y. A., Ling, C. N., Talib, R. A., Taip, F. S., & Aziz, M. G. (2012). Physicochemical and nutritional properties of spray-dried pitaya fruit powder as natural colorant. *Food Science and Biotechnology*, 21, 675–682. https://doi.org/10.1007/s10068-012-0088-z
- Tze, N.L., Han, C.P., Yusof, Y.A., Ling, C.N., Talib, R.A., Taip, F.S., Aziz, M.G. Physicochemical and nutritional properties of spray-dried pitaya fruit powder as natural colorant. *Food Sci. Biotechnol.* 2012, 21, 675–682.
- Udeh, O. (2013). Role of magnesium ions on yeast performance during very high gravity fermentation. *Journal of Brewing* and *Distilling*, 4, 19–45. https://doi.org/10.5897/JBD 2013.0041
- Utpott, M., Ramos, R., Carolina, D. A., Vargas, G., Raisa, A., Paiva, N., Tischer, B., Oliveira, A., & Flôres, S. H. (2020). Characterization and application of red pitaya (*Hylocereus polyrhizus*) peel powder as a fat replacer in ice cream. *Journal of Food Processing and Preservation*, 44(5), 1–10. https://doi.org/10.1111/jfpp.14420
- Van Hoed, V., De Clercq, N., Echim, C., Andjelkovic, M., Leber, E., Dewettinck, K., & Verhé, R. (2009). Berry seeds: A source of specialty oils with high content of bioactives and nutritional value. *Journal of Food Lipids*, 16, 33–49.
- Weller, S.A. Accounting for skill shortages? Migration and the Australian labour market. *Pop. Space Place* 2017, 23, e1997.
- Wichienchot, S., Jatupornpipat, M., & Rastall, R. A. (2010). Oligosaccharides of pitaya (dragon fruit) flesh and their prebiotic properties. *Food Chemistry*, 120, 850–857. https://doi.org/10.1016/j.foodchem.2009.11.026

Wichienchot, S., Jatupornpipat, M., Rastall, R. Oligosaccharides of pitaya (dragon fruit) flesh and their prebiotic properties. Food Chem. 2010, 120, 850–857.

- Wong, Y. M., & Siow, L. F. (2015). Effects of heat, pH, antioxidant, agitation and light on betacyanin stability using red-fleshed dragon fruit (*Hylocereus polyrhizus*) juice and concentrate as models. *Journal of Food Science and Technology*, 52, 3086–3092. https://doi.org/10.1007/s13197-014-1362-2
- Woo, K. K., Chong, Y. Y., Li Hiong, S. K., & Tang, P. Y. (2010). Pectin extraction and characterization from red dragon fruit (*Hylocereus polyrhizus*): A preliminary study. *Journal of Biological Sciences*, 10, 631–636. https://doi. org/10.3923/jbs.2010.631.636
- Wu, L., Hsu, H. W., Chen, Y. C., Chiu, C. C., Lin, Y. I., & Ho, J. A. (2006). Antioxidant and antiproliferative activities of red pitaya. *Food Chemistry*, 95, 319–327. https://doi.org/10.1016/j.foodchem.2005.01.002
- Xu, J., Wang, W., Zhao, Y. Phenolic compounds in whole grain sorghum and their health benefits. *Foods* 2021, *10*, 1921.
- Yadav, A., Garg, S., Kumar, S., Alam, B., Arunachalam, A. A review on genetic resources, breeding status and strategies of dragon fruit. *Genet. Resour. Crop Evol.* 2024, 1–21.
- Yan, Y., Zhou, X., Guo, K., Zhou, F., Yang, H. Use of chlorogenic acid against diabetes mellitus and its complications. J. Immunol. Res. 2020, 2020, 9680508.
- Yee, K., Caryn, C., Yien, K., Ong, Y., Yin, Y., Renee, T., & Hong, L. (2018). Studies on the storage stability of fermented red dragon fruit (*Hylocereus polyrhizus*) drink. *Food Science and Biotechnology*, 27, 1411–1417. https://doi.org/10.1007/s10068-018-0367-4
- Yeh, W.-J., Tsai, C.-C., Ko, J., Yang, H.-Y. Hylocereus polyrhizus peel extract retards alcoholic liver disease progression by modulating oxidative stress and inflammatory responses in C57BL/6 mice. Nutrients 2020, 12, 3884.
- Yien Ong, Y., Siang Tan, W., Rosfarizan, M., Chan, E. S., & Ti Tey, B. (2012). Isolation and identification of lactic acid bacteria from fermented red dragon fruit juices. *Journal of Food Science*, 77, M560–M564. https://doi.org/10.1111/j .1750 -3841.2012.02894.x
- Yong, Y.Y., Dykes, G., Lee, S.M., Choo, W.S. Comparative study of betacyanin profile and antimicrobial activity of red pitahaya (*Hylocereus polyrhizus*) and red spinach (*Amaranthus dubius*). *Plant Foods Hum. Nutr.* 2017, 72, 41–47
- Younis, I.Y., Ibrahim, R.M., El-Halawany, A.M., Hegazy, M.-E.F., Efferth, T., Mohsen, E. Chemometric discrimination of *Hylocereus undulatus* from different geographical origins via their metabolic profiling and antidiabetic activity. *Food Chem.* 2023, 404, 134650.
- Yusof, Y. A., Mohd Salleh, F. S., Chin, N. L., & Talib, R. A. (2012). The drying and tabletting of pitaya powder. *Journal of Food Process Engineering*, 35, 763–771. https://doi.org/10.1111/j.1745-4530.2010.00625.x
- Zaidel, D., Rashid, J., Hazirah, N., Salleh, L., & Kassim, A. (2017). Extraction and characterisation of pectin from dragon fruit (*Hylocereus polyrhizus*) peels. *Chemical Engineering Transactions*, 56, 805–810. https://doi.org/10.3303/CET1756135
- Zee, F., Yen, C., Nishina, M., & Rica, C. (2004). Pitaya (dragon fruit, strawberry pear). *Fruits and Nuts*, University of Hawaii. 9, 1–3.

- Zhuang, Y., Zhang, Y., & Sun, L. (2012). Characteristics of fibre-rich powder and antioxidant activity of pitaya (*Hylocereus undatus*) peels: Characteristics of fibre-rich powder. *International Journal of Food Science & Technology*, 47, 1279–1285. https://doi.org/10.1111/j. 1365-2621.2012.02971.x
- Zulkifli, S.A., Abd Gani, S.S., Zaidan, U.H., Halmi, M.I.E. Optimization of total phenolic and flavonoid contents of
- defatted pitaya (*Hylocereus polyrhizus*) seed extract and its antioxidant properties. *Molecules* 2020, 25, 787.
- Chen, Siyuan, Xu, Cheng-Yuan, Mazhar, Muhammad, Naiker, Mani, 2024. Nutritional Value and Therapeutic Benefits of Dragon Fruit: A Comprehensive Review with Implications for Establishing Australian Industry Standards. VL 29, DOI 10.3390/molecules29235676